

Nanoscale 1,3,5,7-Tetrasubstituted Adamantanes and *p*-Substituted Tetraphenylmethanes for AFM Applications

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Supporting Information

Experimental procedures and AFM conditions, ¹H NMR spectra for **5**, **7**, **8**, **10–20**, ¹³C NMR spectra for **5**, **10–20**.

General methods. Reagent grade solvents were used without further purification. All reagents were purchased from Aldrich Chemical Co. or Lancaster Co. and were used as received. Tetrahydrofuran (99.9%, anhydrous, inhibitor free) and triethylamine (99.5%) were purchased in Aldrich Sure/Seal™ bottles and were well deoxygenated with argon or nitrogen before use in Sonogashira coupling reactions. ¹H and ¹³C NMR spectra were recorded in CDCl₃. Chemical shifts are in δ units (ppm) referenced to residual proton signals in the deuterated solvents. Coupling constants (*J*) are reported in Hertz (Hz). NMR splitting patterns are designated as s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; and br, broad. Brine refers to a saturated aqueous solution of NaCl. Column chromatography was carried out on Davisil silica gel (60–200 mesh). Analytical TLC was performed on Kieselgel 60 F₂₅₄ glass plates. Spots were rendered visible by exposing the plate to UV light. Radial chromatography (RC) was performed on a Chromatotron 7924T using 1, 2 and 4 mm disks covered with silica gel 60PF₂₅₄ containing gypsum. Mass spectra were recorded on an Agilent 1100 Series LC/MSD. High resolution, MALDI and FAB mass spectra were measured by Dr. Brian Arbogast at Oregon State University. Elemental analyses were performed by Robertson Microlit Laboratories, Inc. Madison, NJ.

1,3,5,7-Tetraphenyladamantane (3). 1-Bromoadamantane (3.00 g, 13.9 mmol) was dissolved in benzene (30 mL). Then *t*-BuBr (3.2 mL, 27.9 mmol) and AlCl₃ (0.16 g, 0.12 mmol) was added. The mixture was refluxed for 2 h. The mixture was cooled to rt, filtered, and the residue was washed with benzene (20 mL), water (20 mL) and CHCl₃ (50 mL). The residue was dried and then purified by washing with CHCl₃ in a Soxhlet apparatus for 24 h. The insoluble residue was dried, affording pure 1,3,5,7-tetraphenyladamantane as a white solid (4.70 g, 77%), mp >250 °C (lit.¹ mp 417-419 °C). HR EI MS Calcd for C₃₄H₃₂ 440.25040, found 440.25008.

1,3,5,7-Tetrakis-(4-iodophenyl)adamantane (5). Iodine (2.30 g, 9.06 mmol) was added to a suspension of 1,3,5,7-tetraphenyladamantane (**3**) (2.00 g, 4.54 mmol) in CHCl₃ (50 mL). The mixture was stirred until all iodine dissolved. Then [bis(trifluoroacetoxy)iodo]benzene (**4**)² (3.90 g, 9.06 mmol) was added and the resulting suspension was flushed with N₂ and stirred for 24 h. The suspension was filtered to remove a pink solid. The CHCl₃ solution was washed with 5% aqueous NaHSO₃ (50 mL), water (50 mL) and brine (50 mL). The solution was dried over anhydrous MgSO₄ and concentrated to dryness giving crude **5**, which was crystallized from 9:1 CHCl₃/MeOH yielding tetraiodide **5** (2.34 g, 46%) as colorless crystals, which were pure by NMR and TLC: mp 320-322 °C (lit.¹ 345 °C), R_f = 0.5 (1:1 CHCl₃/hexanes); ¹H NMR δ 2.06 (s, 12H), 7.18 (d, *J* = 8.6 Hz, 8H), 7.67 (d, *J* = 8.6 Hz, 8H); ¹³C NMR δ 39.30, 46.94, 91.99, 127.38, 137.78, 148.66; MS Calcd for C₃₄H₂₈I₄ (M⁺) 943.8, found 943.9. Anal. Calcd for C₃₄H₂₈I₄: C, 43.25; H, 2.99. Found: C, 43.52; H, 2.89.

4-Trityl-iodobenzene (7). 4-Tritylaniline (2.50 g, 7.45 mmol) was added to acetone (70 mL). It dissolved upon addition of concentrated HCl (7 mL) in water (10 mL). The solution was cooled to 0 °C and then sodium nitrite (0.800 g, 11.6 mmol) in water (5 mL) was added dropwise with stirring. The solution was stirred at 0 °C for another 30 min and then potassium iodide (2.00 g, 12.1 mmol) in water (7 mL) was added dropwise. The resulting solution was stirred at 0 °C for 1 h, then at rt for 1h and then at 60 °C for 2 h. Sodium bisulfite (1 g) was added to consume any iodine that may have formed in the

reaction. The mixture was extracted with ether (150 mL). The extract was washed with water (60 mL) followed by brine (2 × 60 mL). The extract dried over anhydrous MgSO₄ and concentrated on the rotary evaporator giving crude **7**. Purification by chromatography on silica gel and elution with 1:1 hexanes/CH₂Cl₂ gave **7** (3.1 g, 94%) as a yellow powder, mp ~180-230 °C (dec. color change to gray), which was pure by TLC and NMR: ¹H NMR(CDCl₃) δ 7.56 (d, *J* = 8.4 Hz, 2H), 7.20 (m, 15H), 6.97 (d, *J* = 8.4 Hz, 2H); MS Calcd for C₂₅H₁₈I (M-1) 445.1, found 445.2.

Tetra(4-iodophenyl)methane (8). A mixture of 4-trityl-iodobenzene **7** (1.29 g, 2.90 mmol), [bis(trifluoroacetoxy) iodo]benzene (2.24 g, 5.21 mmol) and iodine (1.10 g, 4.34 mmol) in CHCl₃ (30 mL) was stirred at rt for 3 h and filtered. The filtrate was concentrated to dryness and the residue was triturated with CHCl₃ (20 mL) and filtered. The resulting solid was combined with the solid obtained from the original filtration and then boiled in CHCl₃ (40 mL) for 10 h. The suspended solid was collected by filtration and dried in vacuum to give tetraiodide **8** (0.50 g, 28%) as a colorless solid, mp >250 °C (lit.³ mp not reported), which was pure by TLC and NMR: ¹H NMR(CDCl₃) δ 6.88 (d, *J* = 8.7 Hz, 8H), 7.58 (d, *J* = 8.7 Hz, 8H); MS Calcd for C₂₅H₁₆I₄ (M⁺) 823.7, found, 823.6.

***N,N*-Bis(2-hydroxyethyl)-4-iodoaniline (10).** Ethylene oxide (12 mL) was condensed at -78 °C and added to *p*-iodoaniline (7.00 g, 32.0 mmol) in methanol (10 mL) at 0 °C in a thick walled reaction tube. The tube was sealed with a Teflon screw cap and stirred at 55 °C for 24 h. The mixture was allowed to cool to rt, and the solvent was evaporated. The residue was chromatographed on silica gel with 1-10% MeOH in CHCl₃ as a eluent to give diol **10** (8.80 g, 90%) as colorless crystals, mp 76-78 °C, which were pure by TLC and NMR: ¹H NMR δ 3.53 (t, *J* = 4.8 Hz, 4H), 3.65 (bs, 1H, disappeared with D₂O exchange), 3.80 (t, *J* = 4.9 Hz, 4H), 6.46 (d, *J* = 9.0 Hz, 2H), 7.45 (d, *J* = 9.0 Hz, 2H); ¹³C NMR δ 55.13, 60.35, 77.60, 114.67, 137.72, 147.24; MS Calcd for C₁₀H₁₅INO₂ (M+1)⁺ 308.0, found 308.0. Anal. Calcd for C₁₀H₁₄INO₂: C, 39.11; H, 4.59; N, 4.56. Found: C, 39.21; H, 4.58, N 4.48.

***N,N*-Bis(2-chloroethyl)-4-iodoaniline (11).** A solution of *N,N*-bis(2-hydroxyethyl)-4-iodoaniline **10** (4.0 g, 13 mmol) in pyridine (5 mL) was added dropwise to a stirred solution of phosphorus oxychloride (2.4 mL, 26 mmol) in pyridine (5 mL) at 0 °C. The mixture was stirred at rt for 30 min and then at reflux temperature for 1.5 h. The mixture was allowed to stir overnight at rt. The solvent was evaporated and the residue was mixed with water (50 mL) and extracted with ether (2 × 50 mL). The combined extracts were washed with brine (2 × 50 mL), dried over anhydrous MgSO₄ and evaporated. The residue was purified by chromatography on silica gel with 20% CH₂Cl₂ in hexane as an eluent to afford dichloride **11** (3.4 g, 76%) as colorless crystals, mp 68.5-69.5 °C, which were pure by TLC and NMR: ¹H NMR δ 3.58-3.73 (m, 8H), 6.47 (d, *J* = 9.3 Hz, 2H), 7.50 (d, *J* = 9.0 Hz, 2H); ¹³C NMR δ 40.13, 53.31, 78.70, 114.22, 138.23, 145.68; MS Calcd for C₁₀H₁₃Cl₂IN (M+1)⁺ 343.9, found, 343.9. Anal. Calcd for C₁₀H₁₂Cl₂IN: C, 34.91; H, 3.52; N, 4.07. Found: C, 35.20; H, 3.38; N, 4.01.

***N,N*-Bis(2-thiocyanatoethyl)-4-iodoaniline (12).** A solution of *N,N*-bis(2-chloroethyl)-4-iodoaniline **11** (2.10 g, 6.10 mmol) and potassium thiocyanate (11.85 g, 122 mmol) in ethanol and water (3 mL) was heated with stirring for 72 h at 125 °C. The solvent was evaporated and the residue was mixed with water (80 mL). The mixture was extracted with ether (100 mL) and chloroform (2 × 60 mL). The combined organic extracts were washed with brine (2 × 60 mL) and water (60 mL), dried over anhydrous MgSO₄ and evaporated. The residue was purified by chromatography on silica gel with CHCl₃ as an eluent to give **12** (1.2 g, 55%) as colorless crystals, mp 89.5-91 °C, which were pure by TLC and NMR: ¹H NMR δ 3.11 (t, *J* = 6.9 Hz, 4H), 3.82 (t, *J* = 6.9 Hz, 4H), 6.56 (d, *J* = 9.3 Hz, 2H), 7.56 (d, *J* = 9.3 Hz, 2H); ¹³C NMR δ 30.65, 51.60, 81.05, 111.43, 115.80, 138.61, 144.79; MS Calcd for C₁₂H₁₃IN₃S₂ (M+1)⁺ 389.95, found 389.9. Anal. Calcd for C₁₂H₁₂IN₃S₂: C, 37.02, H, 3.11, N, 10.79. Found: C, 36.94, H, 3.03, N, 10.63.

5-(4-Iodophenyl)-[1,2,5]dithiazepane (13). A. From 12. A solution of *N,N*-bis(2-thiocyanatoethyl)-4-iodoaniline (**12**) (1.1 g, 2.83 mmol) and potassium thiocyanate (10.23 g, 5.66 mmol) in 90% ethanol (30 mL) was heated with stirring at 125 °C for 2 h. The solvent was evaporated and the residue was mixed with water (60 mL). The mixture was extracted with ether (2 × 50 mL) and chloroform (50 mL). The combined organic extracts were washed with brine (2 × 50 mL), dried over anhydrous MgSO₄ and evaporated. The residue was purified by chromatography on silica gel with 1:4 CHCl₃/hexanes as an eluent to give **13** (0.865 g, 91%) as colorless crystals, mp 98-100 °C (dec.).

B. From 11. A suspension of dichloride **11** (6.69 g, 19.4 mmol) and potassium thiocyanate (37.8 g, 389 mmol) in 10:1 ethanol/water (110 mL) was heated with stirring for 36 h in a 125 °C bath. The solvent was removed and the residue was treated with water (60 mL) and extracted with ether (50 mL) and then chloroform (2 × 60 mL). The combined organic extracts were washed with brine (2 × 50 mL) and dried over anhydrous MgSO₄. Evaporation of the solvent gave a brown oil, which was purified by chromatography over a short column of silica gel. Elution with 1:4 CHCl₃/hexanes gave **13** (4.64 g, 71%) as colorless crystals, mp 99-101 °C (from 1:10 CHCl₃/hexanes): R_f = 0.75 (CHCl₃); ¹H NMR δ 3.06 (t, *J* = 5.6 Hz, 4H), 3.93 (t, *J* = 5.6 Hz, 4H), 6.42 (d, *J* = 8.7 Hz, 2H), 7.46 (d, *J* = 8.7 Hz, 2H); ¹³C NMR δ 36.58, 52.25, 77.13, 113.54, 138.01, 145.90; MS Calcd for C₁₀H₁₃INS₂ (M+1)⁺ 337.9, found 338.0. Anal. Calcd for C₁₀H₁₂INS₂: C, 35.61; H, 3.59; N, 4.15. Found: C, 35.79; H, 3.53; N 4.07.

5-(4-Trimethylsilylanylethynylphenyl)-[1,2,5]dithiazepane (14). Iodide **13** (1.00 g, 2.97 mmol), trimethylsilylacetylene (2.91 g, 29.70 mmol), Pd(PPh₃)₂Cl₂ (0.104 g, 0.149 mmol) and CuI (0.056 g, 0.297 mmol) were added to degassed triethylamine (5 mL) and THF (5 mL) in a thick walled reaction tube. The tube was sealed with a Teflon screw cap and the contents were stirred at 70 °C for 24 h. The reaction mixture was filtered and the solid was washed with diethyl ether (60

mL). The organic phases were combined and evaporated to give crude **14**, which was purified by chromatography on silica gel. Elution with 1:4 ether/hexanes gave **14** (0.83 g, 91%) as a yellow crystalline solid, which was pure by TLC and NMR: mp 100-101 °C (from 1:6 Et₂O/hexanes); R_f = 0.4 (1:4 Et₂O/hexanes); ¹H NMR δ 0.23 (s, 9H), 3.06 (t, *J* = 5.6 Hz, 4H), 3.95 (t, *J* = 5.6 Hz, 4H), 6.53 (d, *J* = 9.0 Hz, 2H), 7.34 (d, *J* = 9.0 Hz, 2H); ¹³C NMR 0.17, 36.67, 52.40, 91.46, 106.14, 110.32, 110.94, 133.57, 146.46; MS Calcd for C₂₁H₂₂NS₂Si (M+1) 308.1, found 308.1. Anal. Calcd for C₁₅H₂₁NS₂Si (307.55): C, 58.58; H, 6.88; N, 4.55. Found: C, 58.31; H, 6.84; N, 4.34.

5-(4-Ethynylphenyl)-[1,2,5]dithiazepane (15). To a stirred solution of TMS derivative **14** (1.30 g, 4.23 mmol) in tetrahydrofuran (6 mL) at -20 °C, a 1 M solution of *n*-Bu₄NF in tetrahydrofuran (4.2 mL, 4.2 mmol) was added. After 30 min, the mixture was diluted with water (50 mL) and extracted with CH₂Cl₂ (50 mL) and then ether (50 mL). The combined organic extracts were washed with brine (2 × 50 mL), dried over anhydrous MgSO₄ and evaporated. The residue was purified by chromatograph on silica gel. Elution with 1:9 ether/hexane gave **15** (0.85 g, 86%) as a colorless crystalline solid, which was pure by TLC and NMR: mp 119-121 °C (from 1:1 CHCl₃/hexanes); R_f = 0.25 (1:9 Et₂O/hexanes); ¹H NMR δ 2.97 (s, 1H), 3.07 (t, *J* = 5.6 Hz, 4H), 3.97 (t, *J* = 5.6 Hz, 4H), 6.56 (d, *J* = 9 Hz, 2H), 7.37 (d, *J* = 9.0 Hz, 2H); ¹³C NMR δ 36.64, 52.39, 74.97, 84.49, 109.19, 119.96, 133.66, 146.63; MS Calcd for C₁₂H₁₄NS₂ (M+1)⁺ 236.0, found 236.0. Anal. Calcd for C₁₂H₁₃NS₂: C, 61.23; H, 5.57; N 5.95. Found: C, 61.35; H, 5.53; N 5.95.

5-[4-(4-Trimethylsilanylethynylphenylethynyl)phenyl]-[1,2,5]dithiazepane (16). 5-(4-Ethynylphenyl)-[1,2,5]dithiazepane **15** (0.77g, 3.28 mmol), 4-iodophenylethynyltrimethylsilane (1.08 g, 3.60 mmol), Pd(PPh₃)₂Cl₂ (0.569 g, 0.811 mmol) and CuI (0.308 g, 1.617 mmol) were added to degassed triethylamine (10 mL) and tetrahydrofuran (5 mL) in a thick walled reaction tube. The tube was sealed with a Teflon screw cap and stirred at 40-45 °C for 2 h, then at rt for 14 h. The reaction mixture was filtered and the solid was washed with diethyl ether (50 mL). The combined organic solutions were evaporated. The residue was chromatographed on silica gel with 10% ether in hexane as a eluent to give compound **16** (0.83 g, 91%) as colorless crystals, mp 188-190 °C, which were pure by TLC and NMR: ¹H NMR δ 7.41 (s, 4H), 7.40 (d, *J* = 9.2 Hz, 2H), 6.59 (d, *J* = 9.2 Hz, 2H), 3.98 (t, *J* = 5.6 Hz, 4H), 3.08 (t, *J* = 5.6 Hz, 4H), 0.26 (s, 9H); ¹³C NMR δ 146.51, 133.24, 131.81, 130.02, 124.18, 121.98, 111.14, 110.15, 104.87, 95.76, 92.43, 87.31, 52.45, 36.68, -0.07; MS Calcd for C₂₃H₂₆HS₂Si (M+1) 408.1, found 408.1. Anal. Calcd for C₂₃H₂₅NS₂Si: C, 67.76; H, 6.18; N, 3.44. Found: C, 67.51; H, 5.98; N, 3.45.

5-[4-(4-Ethynylphenylethynyl)phenyl]-[1,2,5]dithiazepane (17). Dithiazepane **16** (0.37g, mmol) was mixed with potassium carbonate (0.5 g, mmol) in CHCl₃ (4 mL) and CH₃OH (12 mL). The suspension was stirred at rt for 3 h and then diluted with ether (100 mL). The organic phase was washed with brine (2 × 30 mL) and water (30 mL), then dried over anhydrous MgSO₄ and evaporated. The residue was purified by chromatograph on silica gel with 10% ether in hexane as the eluent to give **17** (1.098 g, 82%) as yellow crystals, mp 159-161 °C, which were pure by TLC and NMR: ¹H NMR δ 7.41 (s, 4H), 7.40 (d, *J* = 9.15 Hz, 2H), 6.59 (d, *J* = 9.15 Hz, 2H), 3.98 (t, *J* = 5.55 Hz, 4H), 3.08 (t, 4H, *J* = 5.55 Hz), 0.26 (s, 9H); ¹³C NMR δ 146.55, 133.25, 131.97, 131.09, 124.60, 120.92, 111.12, 99.94, 92.51, 87.12, 83.46, 78.50, 52.45, 36.67; MS Calcd for C₂₀H₁₈NS₂ (M+1) 336.1, found 336.1. Anal. Calcd for C₂₀H₁₇NS₂ (335.49): C, 71.60; H, 5.11; N 4.18. Found: C, 71.37; H, 4.96; N, 3.99.

Tetrakis-{4-[4-(5-[1,2,5]dithiazepanyl)phenylethynyl]phenyl}methane (Molecular Tip 18). Tetra(4-iodophenyl)-methane (**8**, 41.2 mg, 0.0500 mmol), 5-(4-ethynylphenyl)-[1,2,5]-dithiazepane **15** (56.4 mg, mmol), Pd(PPh₃)₂Cl₂ (22 mg, 0.033 mmol) and CuI (12.1 mg, 0.062 mmol) were added to degassed triethylamine (8 mL) and tetrahydrofuran (2 mL) in a thick walled reaction tube which was then flushed with N₂ and capped with a self sealing rubber-lined Teflon screw cap. The mixture was stirred at rt for 2 h, then at 50-60 °C for 4 h. The reaction mixture was filtered and the solid was washed with CHCl₃ (30 mL). The filtrate was evaporated to dryness and the residue was purified by chromatography over a short column of silica gel. Elution with CHCl₃ gave crude **18**, which was further purified by radial chromatography (1 mm disk). Sequential elution with 30% chloroform in hexanes, 50% chloroform in hexanes and 80% chloroform in hexanes afforded **18** (21 mg, 21%) as a yellow glass, which was pure by TLC and NMR: R_f = 0.3 (80% CHCl₃ in hexanes); ¹H NMR δ 3.09 (t, *J* = 5.6 Hz, 16H), 3.98 (t, *J* = 5.6 Hz, 16H), 6.59 (d, *J* = 9.2 Hz, 8H), 7.15 (d, *J* = 8.6 Hz, 8H), 7.388 (d, *J* = 9.2 Hz, 8H), 7.394 (d, *J* = 8.6 Hz, 8H); ¹³C NMR δ 36.74, 52.43, 77.20, 87.16, 90.54, 110.10, 110.48, 121.89, 130.63, 130.86, 133.20, 145.40, 146.30. MS Calcd for C₇₃H₆₄N₄S₈ (M+1)⁺ 1253.3, found 1253.3.

Tetrakis-{4-[4-[4-(5-[1,2,5]dithiazepanyl)phenylethynyl]phenyl]ethynylphenyl}methane (Molecular Tip 19). 5-[4-(4-Ethynylphenylethynyl)phenyl]-[1,2,5] dithiazepane (**17**, 50 mg, 1.64 mmol), tetra(4-iodophenyl)methane (**8**, 21 mg, 0.025 mmol), Pd(PPh₃)₂Cl₂ (11 mg, 0.017 mmol) and CuI (6.1 mg, 0.031 mmol) were added to degassed triethylamine (3 mL) and tetrahydrofuran (3 mL) in a thick walled reaction tube. The tube was sealed with a Teflon screw cap and stirred at 100-105 °C for 3 h, then at rt for 12 h. The reaction mixture was filtered and the solid was washed with diethyl ether (25 mL). The combined organic phase was evaporated. The residue was chromatographed on silica gel with 10% hexanes in chloroform as the eluent to give a solid, which was crystallized by vapor diffusion of hexanes into a chloroform solution. Molecular tip **19** (5.8 mg, 15%) was thus obtained as a yellow crystalline solid, mp >250 °C, which was pure by TLC and NMR: ¹H NMR δ 7.46 (s, 16H), 7.45 (d, *J* = 9 Hz, 8H), 7.41 (d, *J* = 9 Hz, 8H), 7.21 (d, *J* = 8.6 Hz, 8H), 6.60 (d, *J* = 8.6 Hz, 8H), 3.99 (t, *J* = 5.5 Hz, 16H), 3.09 (t, *J* = 5.5 Hz, 16H); ¹³C NMR δ 36.71, 52.48, 77.21, 87.40, 89.78, 90.49, 92.41, 110.23,

111.15, 121.26, 122.09, 124.02, 130.91, 131.09, 131.18, 131.49, 133.26, 145.95, 146.51. MS MALDI Calcd for $C_{105}H_{82}N_4S_8$ (M+2) 1654.4, found 1654.4. Anal. Calcd for $C_{105}H_{80}N_4S_8 \cdot H_2O$: C, 75.41; H, 4.94; N, 3.35. Found: C, 75.22; H, 4.67; N, 3.18.

Tetrakis{4-(4-[5-[1,2,5]dithiazepanyl)phenyl]ethynyl)phenyl}adamantane (Molecular Tip 20). 5-(4-Ethynylphenyl)-[1,2,5]dithiazepane **15** (51 mg, 0.22 mmol), tetra-(4-iodophenyl)adamantane (**5**, 40 mg, 0.042 mmol), $Pd(PPh_3)_2Cl_2$ (6 mg, 0.09 mmol) and CuI (3.4 mg, 0.018 mmol) were added to degassed triethylamine (8 mL) and tetrahydrofuran (2 mL) in a thick walled reaction tube. The tube was sealed with a Teflon screw cap and stirred at 80-85 °C for 5 h, then at rt for 12 h. The reaction mixture was filtered and the solid was washed with $CHCl_3$ (30 mL). The filtrate was evaporated to dryness and the residue was purified by chromatography on silica gel with 70% chloroform in hexanes as the eluent to give **20** (6.8 mg, 12%) as a yellow glass, which was pure by TLC and NMR: $R_f = 0.8$ ($CHCl_3$); 1H NMR δ 2.15 (s, 12 H), 3.09 (t, $J = 5.5$ Hz, 16H), 3.98 (t, $J = 5.5$ Hz, 16H), 7.39-7.50 (m, 24H); ^{13}C NMR δ 36.76, 39.27, 46.90, 52.44, 87.39, 90.01, 110.63, 111.12, 121.90, 125.01 131.35, 133.18, 146.26, 148.60; MS FAB Calcd for $C_{82}H_{77}N_4S_8$ 1373.4 (M+1), found 1373.5.

Atomic Force Microscopy Experiments. AFM measurements were carried out with a Nanoscope IIIa Multimode AFM (Digital Instruments, Santa Barbara, CA) using a 10 μm scanner. Tapping mode AFM (TMAFM) scans were performed in air with NANOSENSORS silicon cantilevers/tips: type NCH, cantilever resonance frequency $f_0 = 289$ -332 kHz and force constant 24.0-37.0 N/m.

The instrument was operated at frequencies slightly lower than the natural resonance frequency in air. All data were recorded in height mode. Set point values were chosen so that the interaction of the tip and sample provided a good compromise between stability and resolution, without damaging the tip or the sample. Scan rates ranged from 1 to 3 Hz. Images were taken at a 512×512 -pixel resolution to increase the detail in the images. All TMAFM studies were carried out on freshly cleaved muscovite mica, grade V-4 (Structure Probe, Inc.). The average sizes of height and width of nanometer sized features and their standard deviation were determined by counting the features on at least three different areas of the same sample and taking the average.

Sample Preparation. Spectroscopic grade solvents were used for sample preparation. *Molecular tip 19 on mica.* One drop of a 0.06 μM , 0.25 μM or 1 μM solution of molecular tip **19** in CH_2Cl_2 was spin-coated onto freshly cleaved mica (cleaved with Scotch tape) for 15-20 s under ambient conditions at a speed of 2000 rpm. Scanning of the sample by AFM began immediately after completion of the spin-coating step.

DNA/molecular tip 19 on mica. λ DNA *Hind* III digest fragments were purchased from New England BioLabs and used without additional purification. Prior to imaging, the DNA was diluted to a concentration of 0.33 $\mu g/mL$ in HEPES buffer (pH = 7.4), containing 4 mM HEPES, 10 mM NaCl, 2 mM $MgCl_2$. An aliquot (20 μL) of the solution was deposited onto freshly cleaved mica. The sample was incubated for 10-15 min in air. The sample was then rinsed well with deionized water, blotted at the edges and allowed to dry under a flow of argon. Next, a 0.25 μM solution of molecular tip **19** in CH_2Cl_2 was spin-coated over the DNA on mica for 15-20 s under ambient conditions at a speed of 2000 rpm. Scanning of the sample by AFM began immediately after completion of the spin-coating step.

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(3) Su, D.; Menger, F. M. *Tetrahedron Lett.* **1997**, 38, 1485-1488.

quan11020130a

Pulse Sequence: s2pul

Solvent: CDCl3

Ambient temperature

INOVA-300 "sunofnar"

PULSE SEQUENCE

Relax. delay 1.000 sec

Pulse 37.9 degrees

Acq. time 2.501 sec

Width 4799.0 Hz

8 repetitions

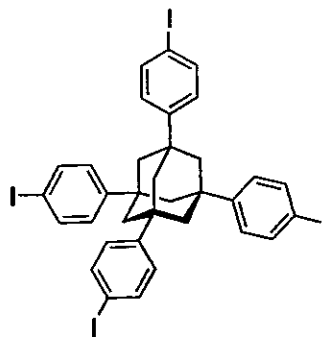
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DATA PROCESSING

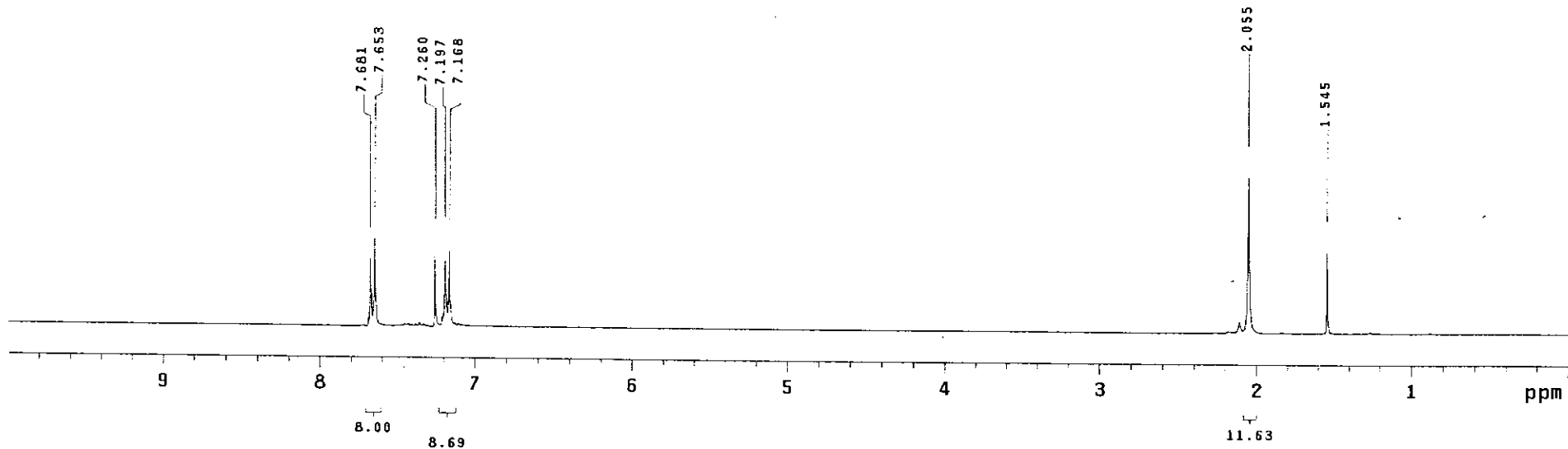
Line broadening 0.2 Hz

FT size 32768

Total time 0 min, 28 sec

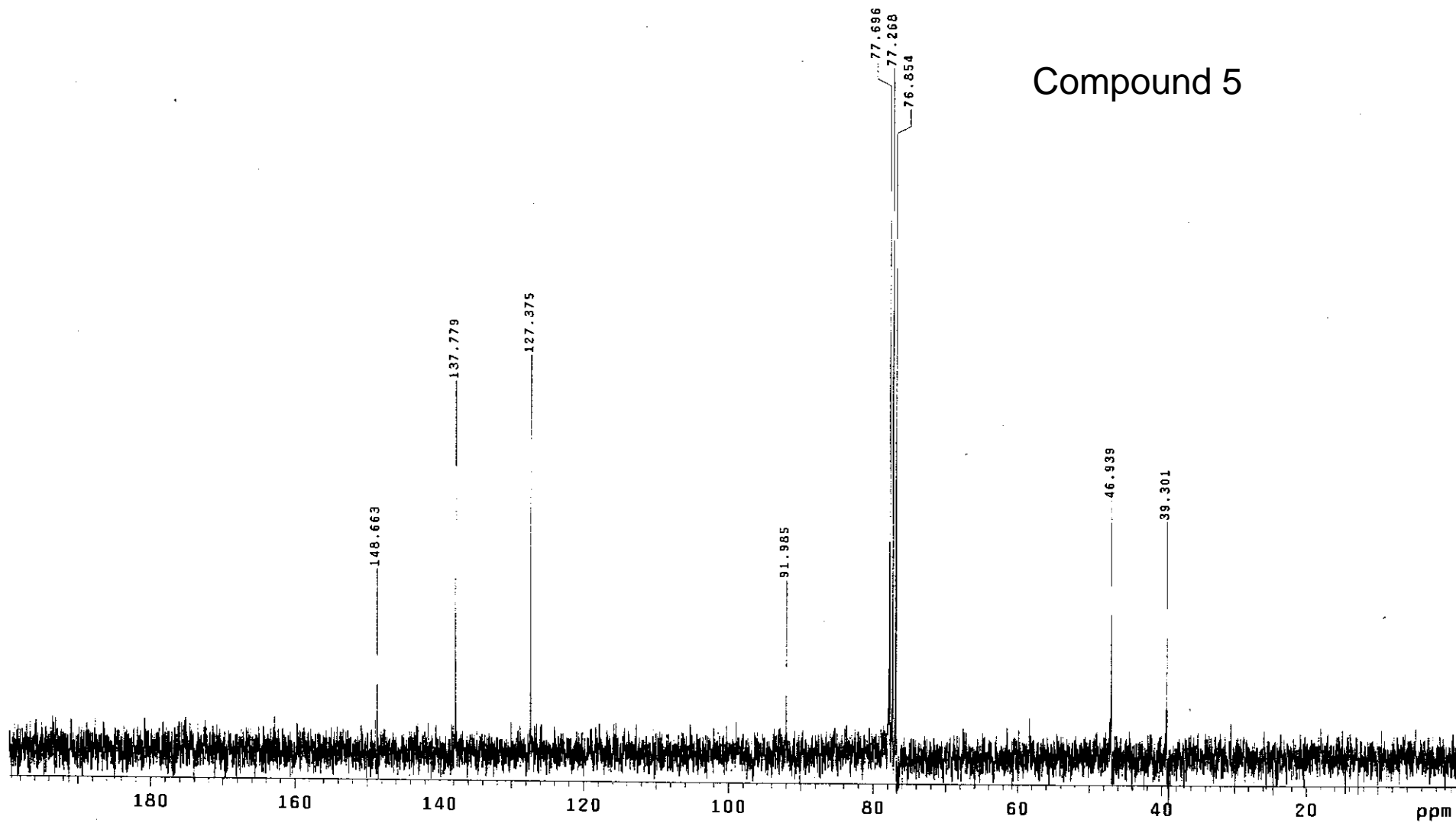


5



quanli020130b

Pulse Sequence: s2pu1



Compound 5

quanli011211b

Pulse Sequence: s2pu1

Solvent: CDCl3

Ambient temperature

INOVA-300 "sunofmr"

PULSE SEQUENCE

Relax. delay 1.000 sec

Pulse 37.9 degrees

Acq. time 2.501 sec

Width 4799.0 Hz

8 repetitions

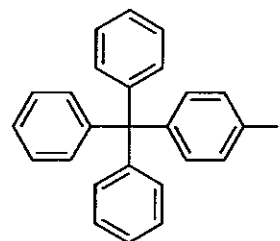
OBSERVE H1, 299.9468612 MHz

DATA PROCESSING

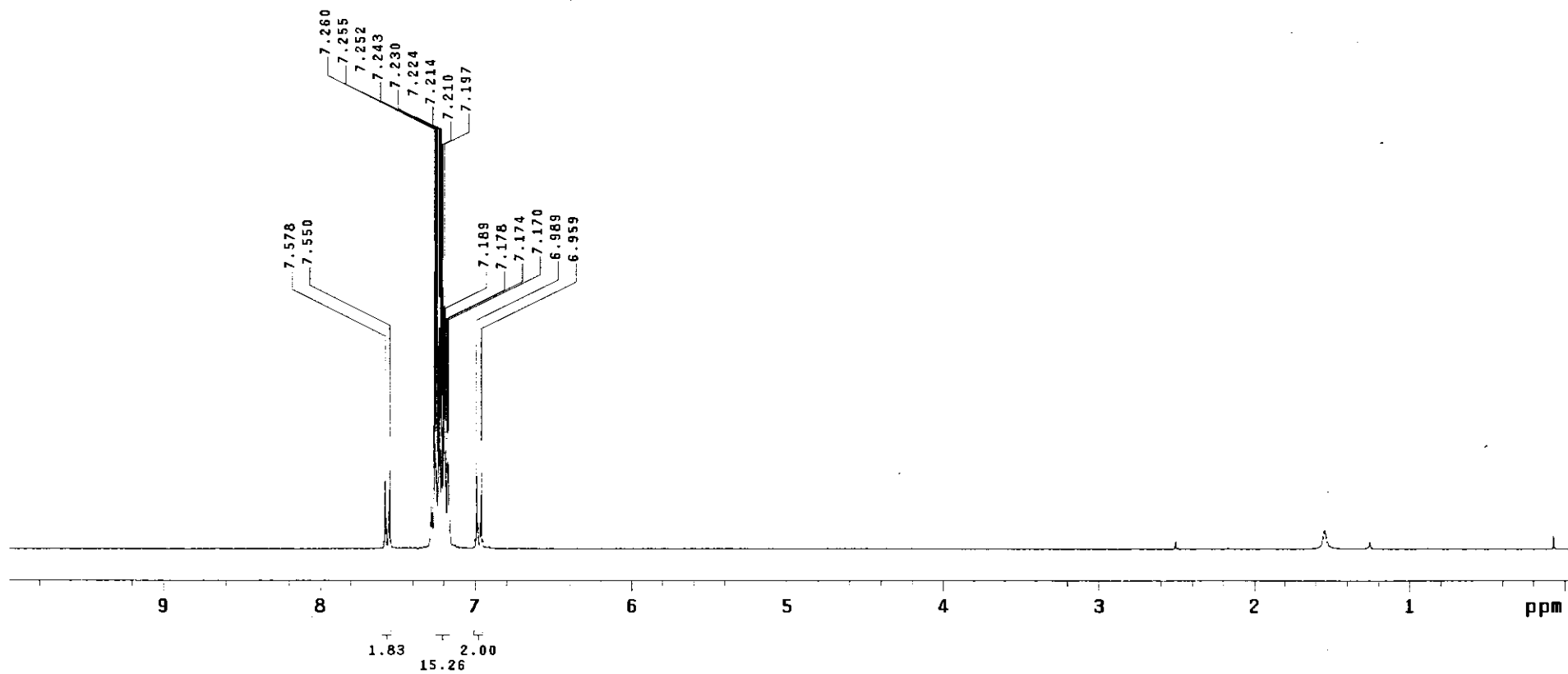
Line broadening 0.2 Hz

FT size 32768

Total time 0 min, 28 sec



7



quan11011226a

Pulse Sequence: s2pu1

Solvent: CDC13

Ambient temperature

File: q11011226a

INOVA-300 "sunofmr"

PULSE SEQUENCE

Relax. delay 1.000 sec

Pulse 37.9 degrees

Acq. time 2.501 sec

Width 4799.0 Hz

8 repetitions

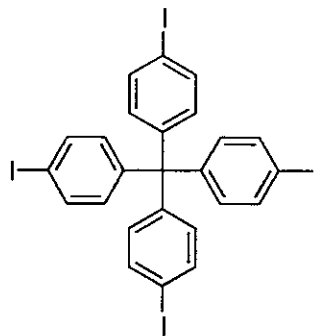
OBSERVE H1, 299.9468609 MHz

DATA PROCESSING

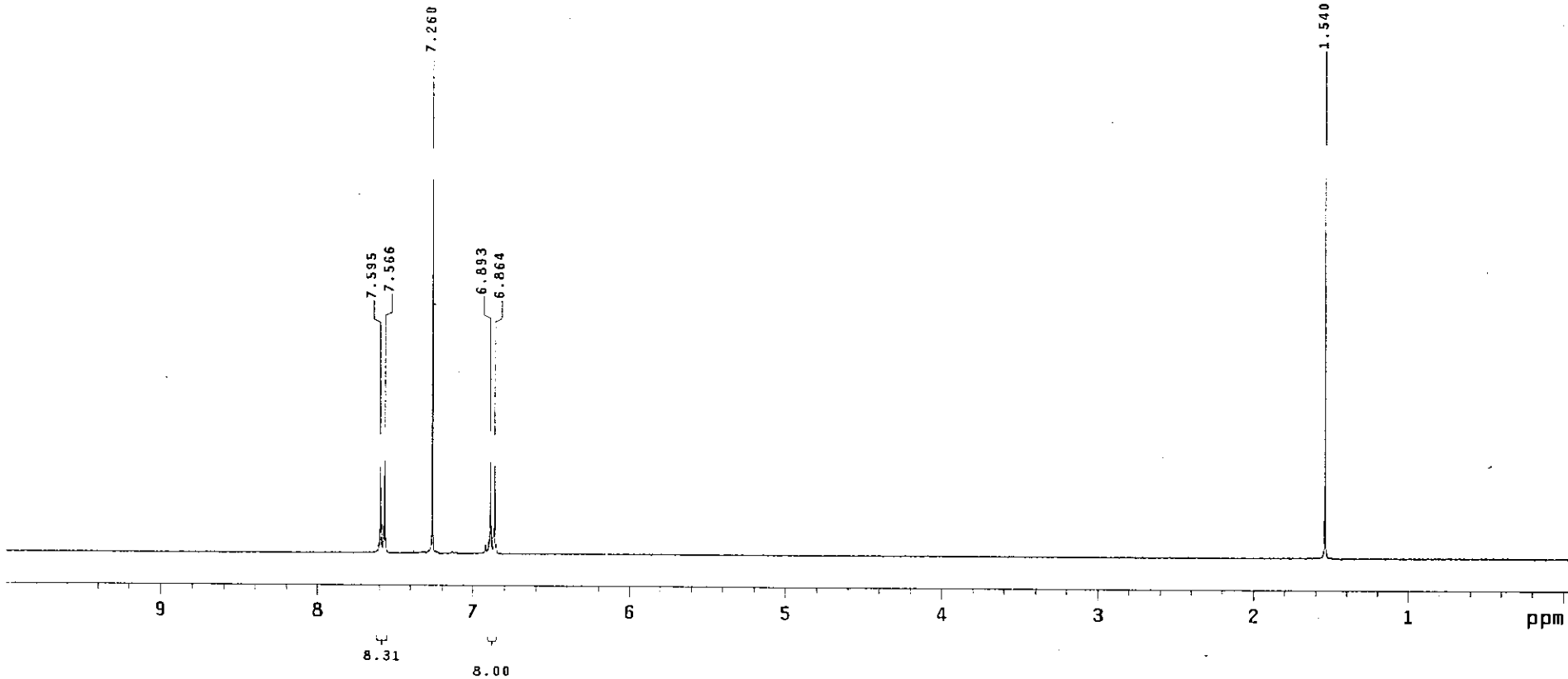
Line broadening 0.2 Hz

FT size 32768

Total time 0 min, 28 sec

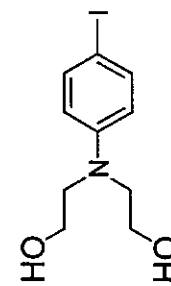


8

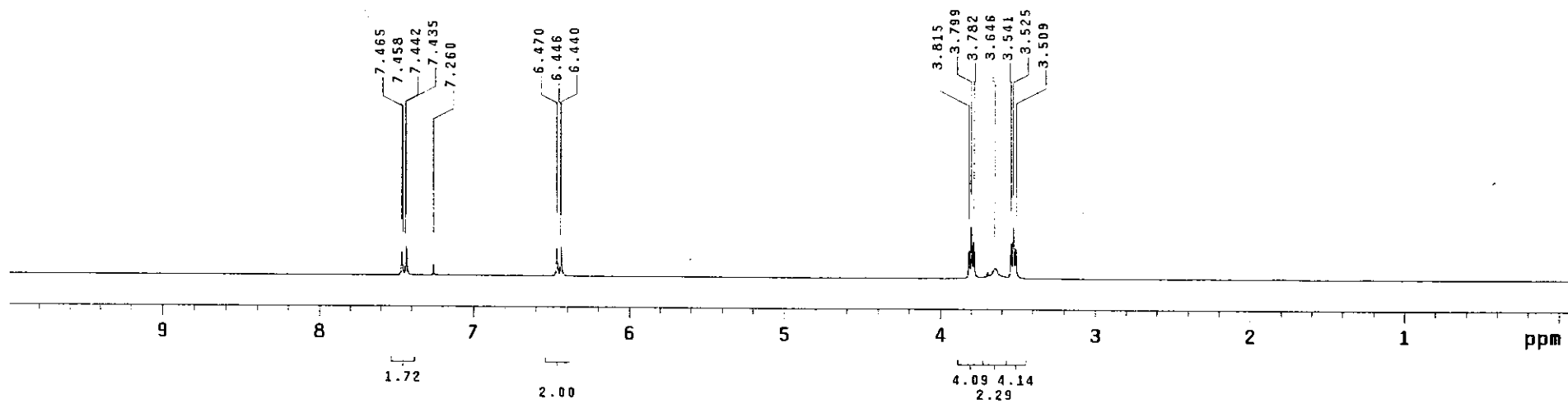


STANDARD 1H OBSERVE

Pulse Sequence: s2pu1



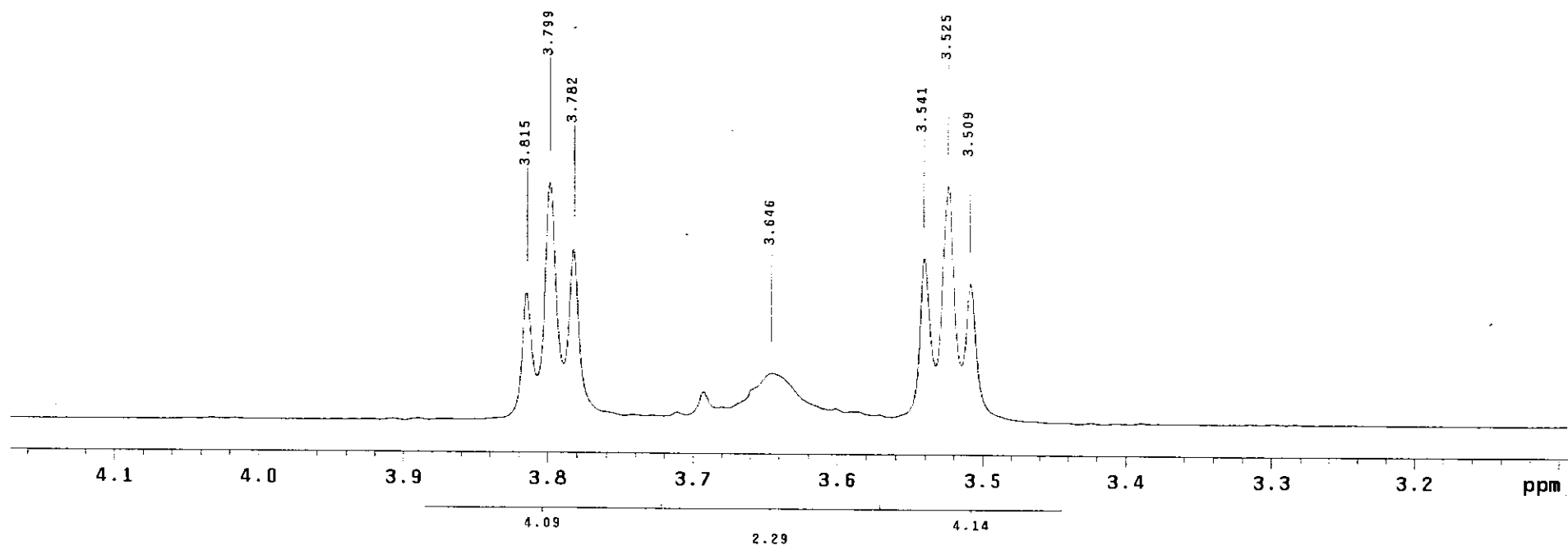
10



STANDARD 1H OBSERVE

Pulse Sequence: s2pul

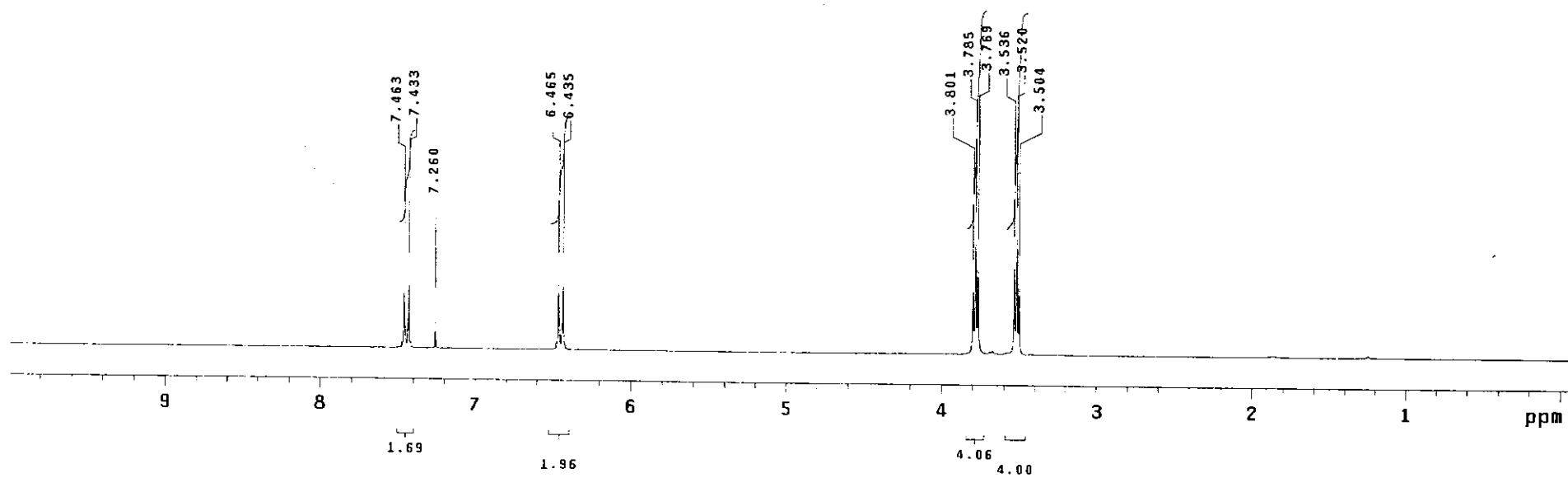
Compound 10



q11011017 D2O exchange

Pulse Sequence: s2pu1

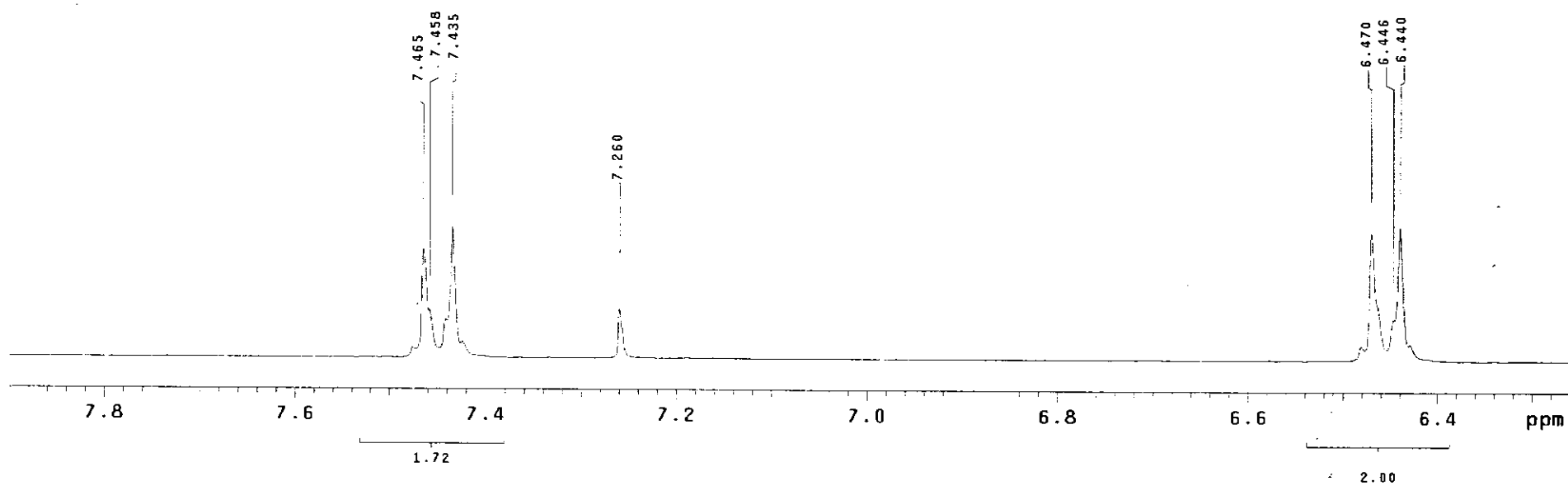
Compound 10



STANDARD 1H OBSERVE

Pulse Sequence: s2pu1

Compound 10



Compound10

Pulse Sequence: s2pu1

Solvent: CDCl3
Ambient temperature
INOVA-300 "sunofmr"

PULSE SEQUENCE

Relax. delay 1.000 sec
Pulse 42.6 degrees
Acq. time 0.599 sec
Width 16501.7 Hz
80 repetitions

OBSERVE C13, 75.4217031 MHz

DECOUPLE H1, 299.9478514 MHz

Power 32 dB
continuously on
WALTZ-16 modulated

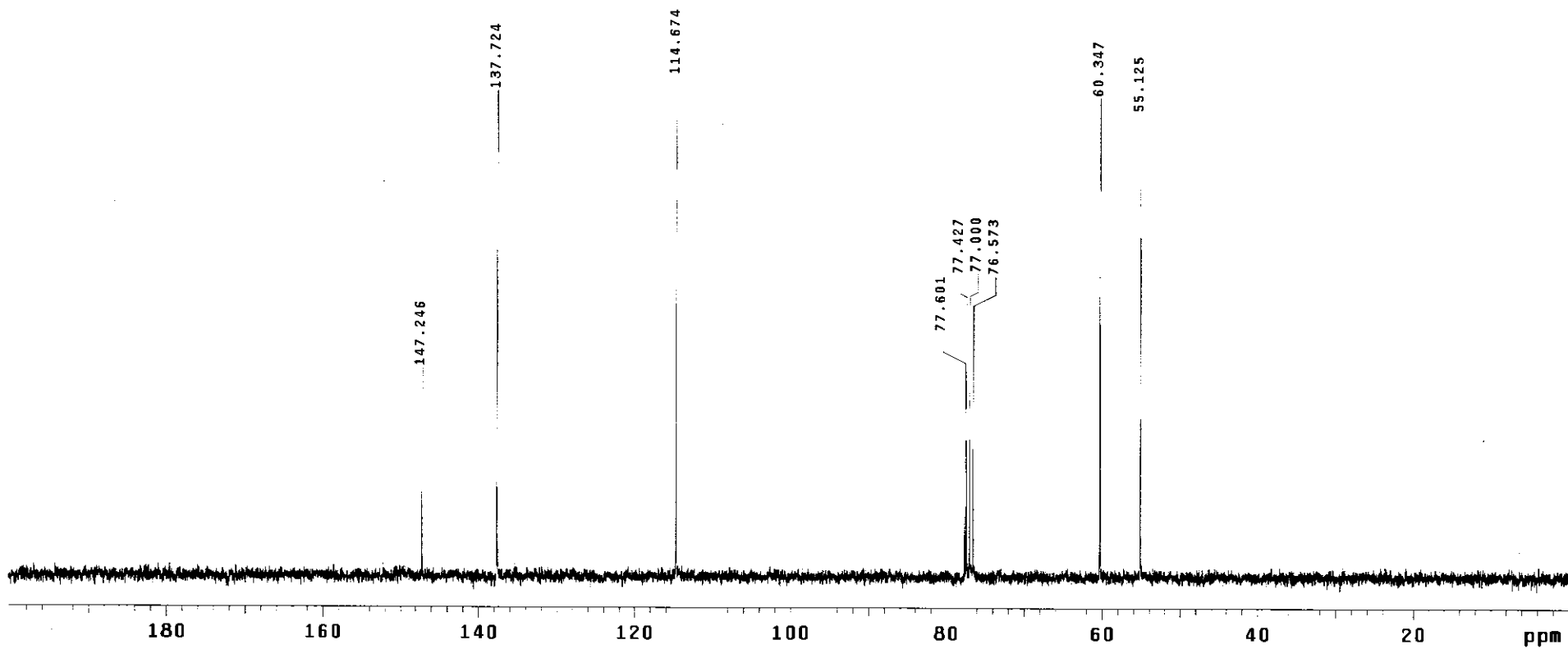
DATA PROCESSING

Line broadening 1.0 Hz

FT size 32768

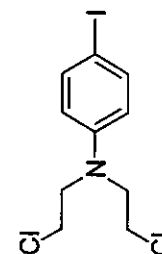
Total time 8 min, 2 sec

Compound 10

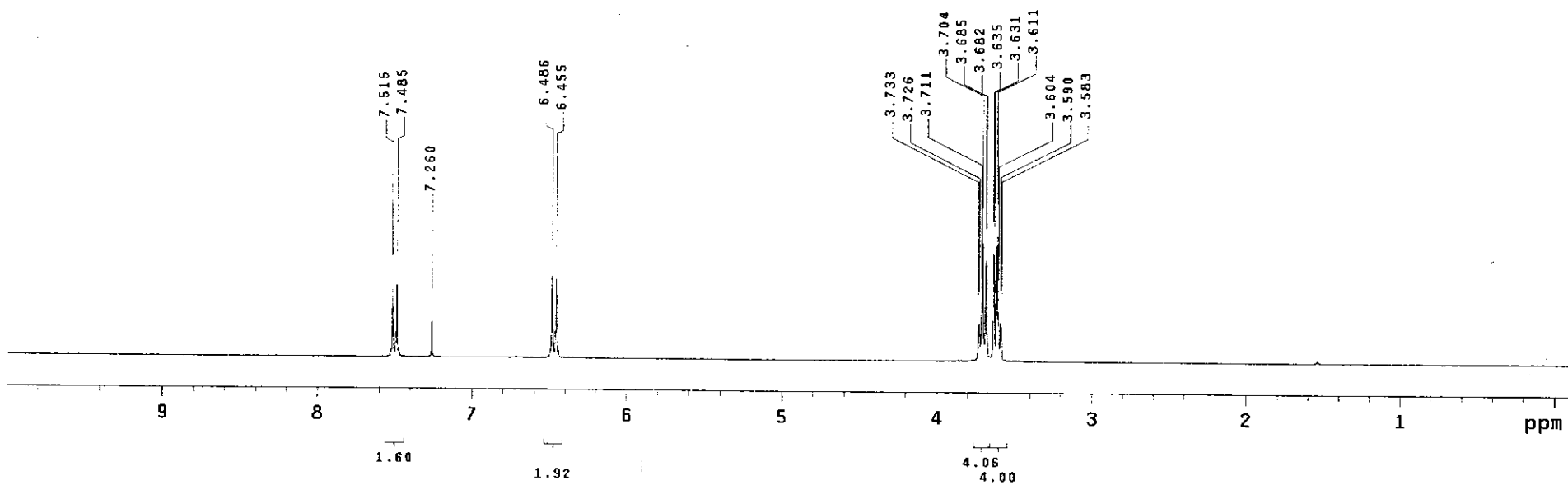


q11011022a

Pulse Sequence: s2pu1



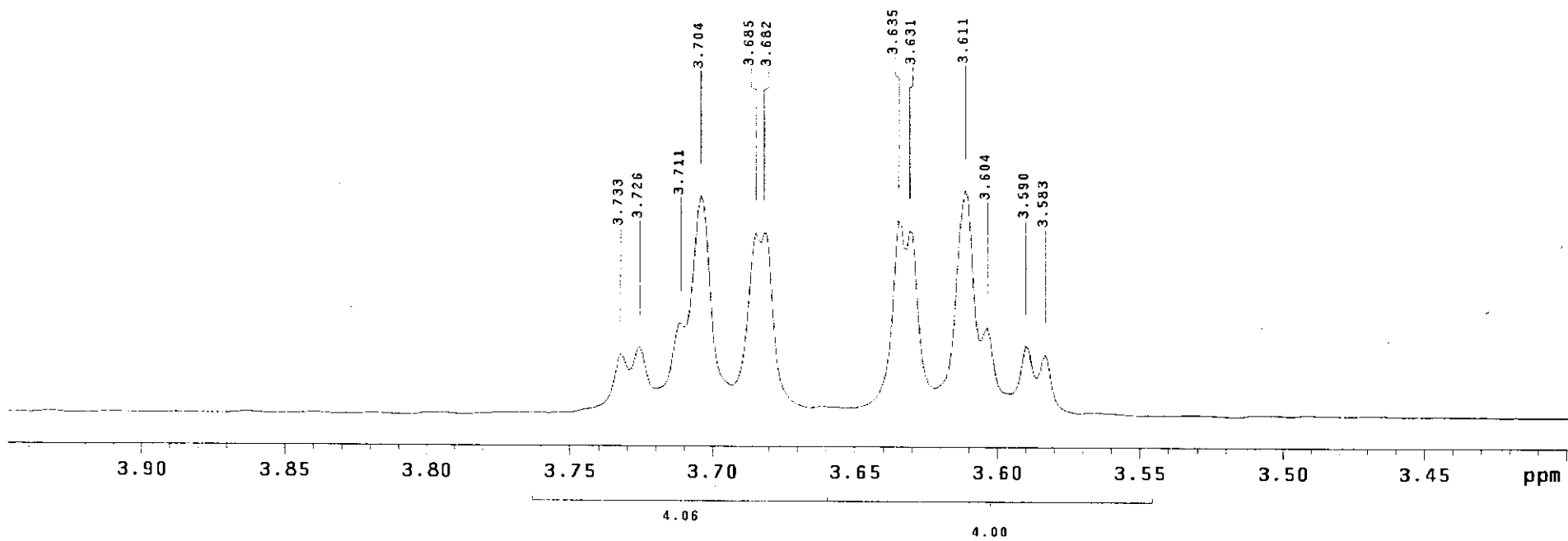
11



q1i011022a

Pulse Sequence: s2pu1

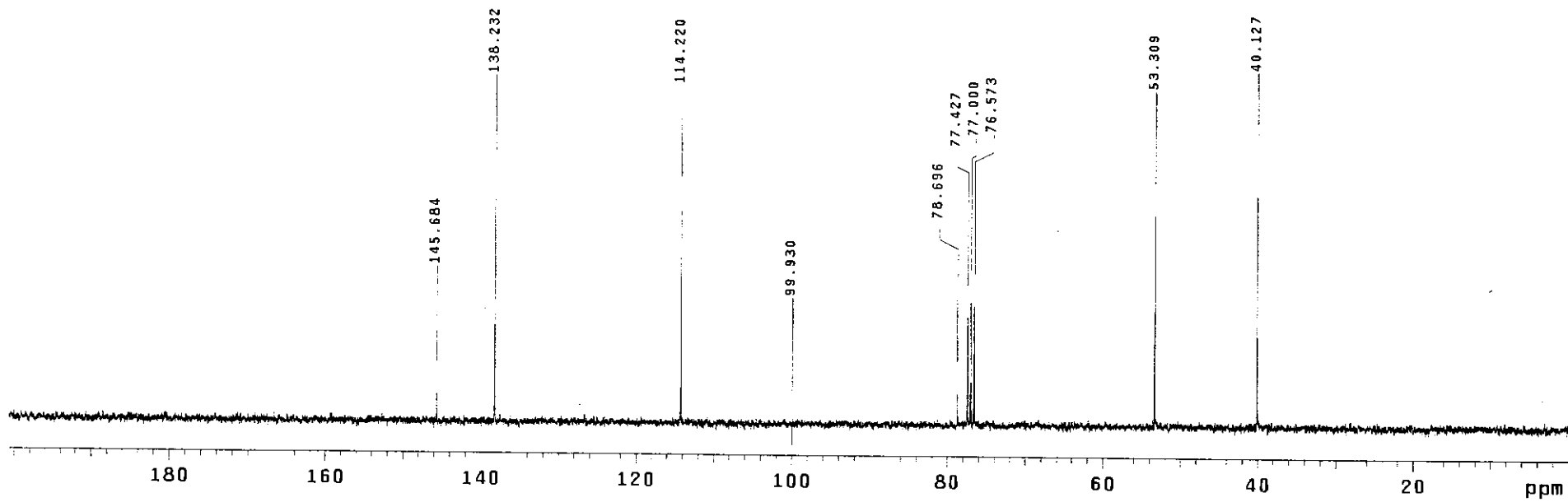
Compound 11



q1i011022d

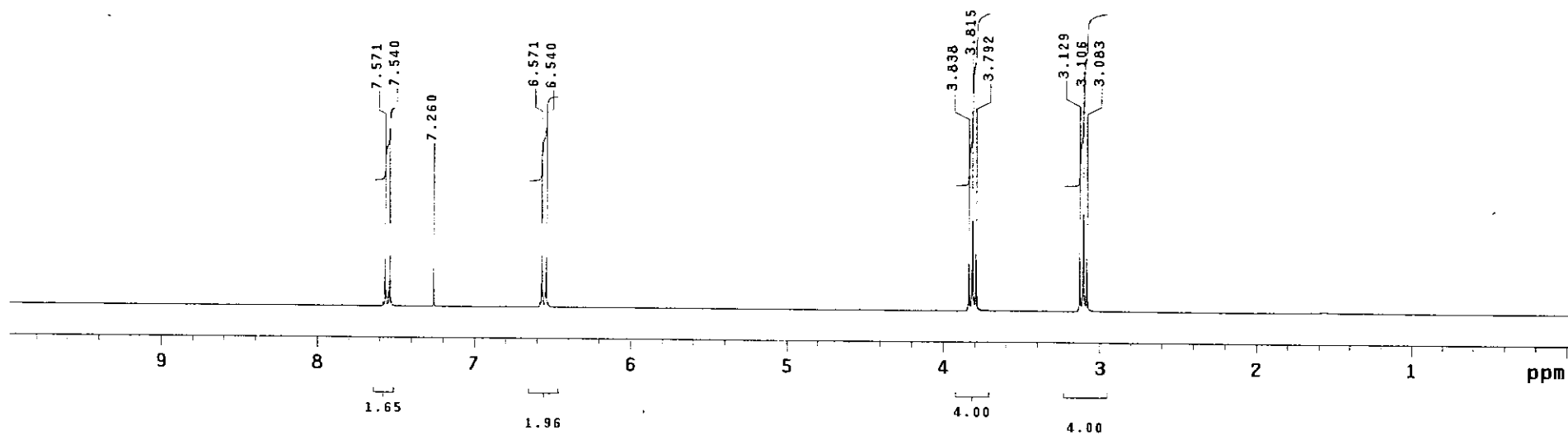
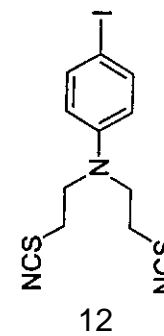
Pulse Sequence: s2pu1

Compound 11



ql1011030a

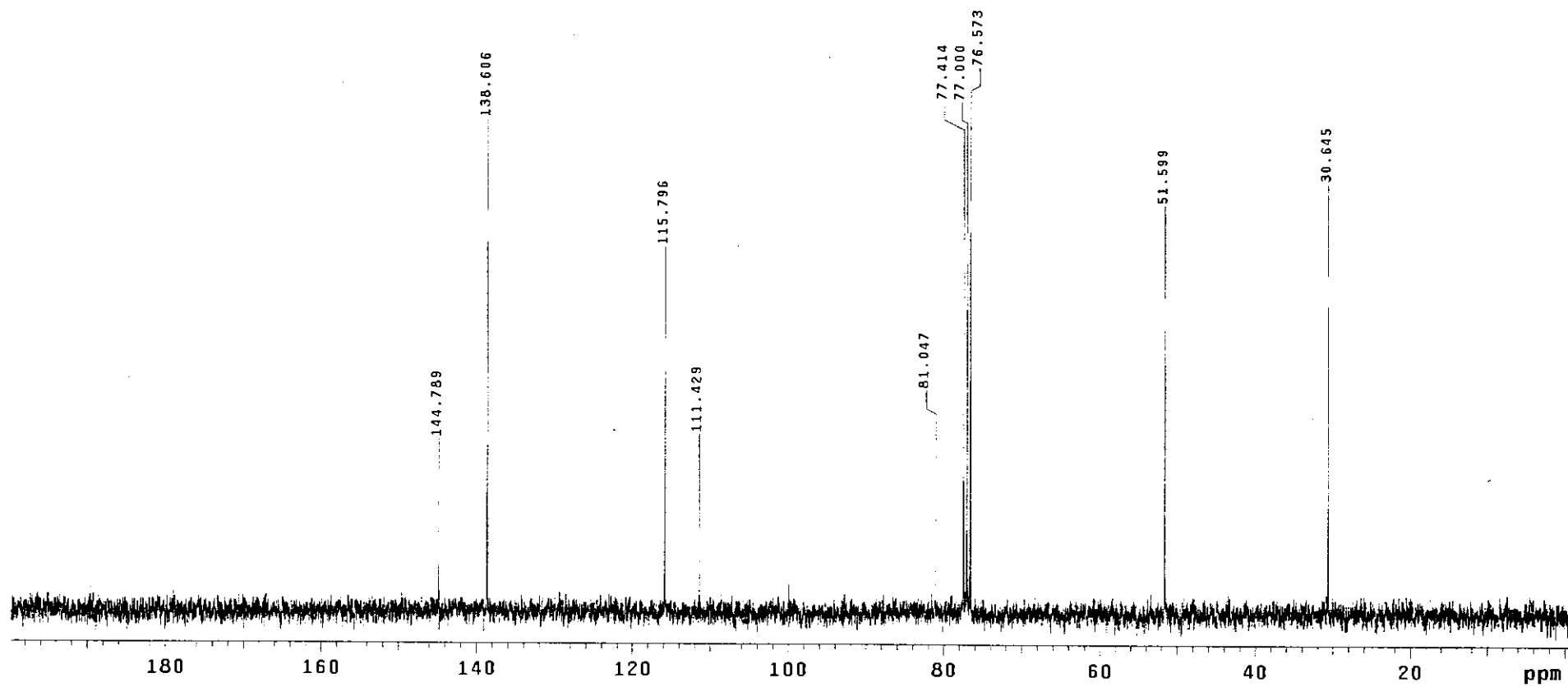
Pulse Sequence: s2pu1



q1i011030b

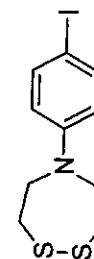
Pulse Sequence: s2pu1

Compound 12

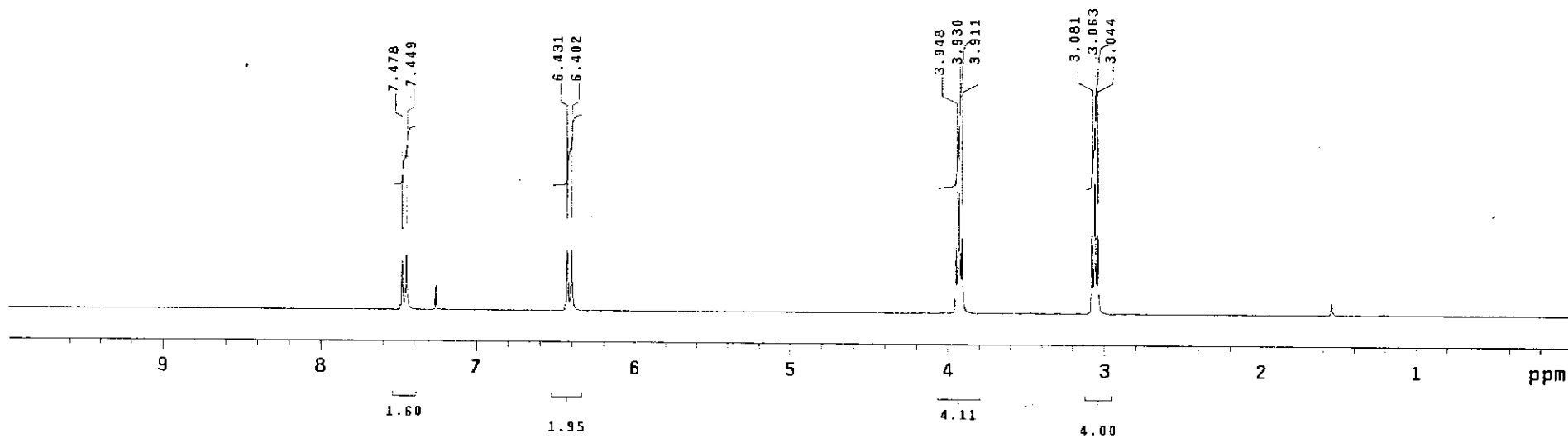


q11011106a

Pulse Sequence: szpu1



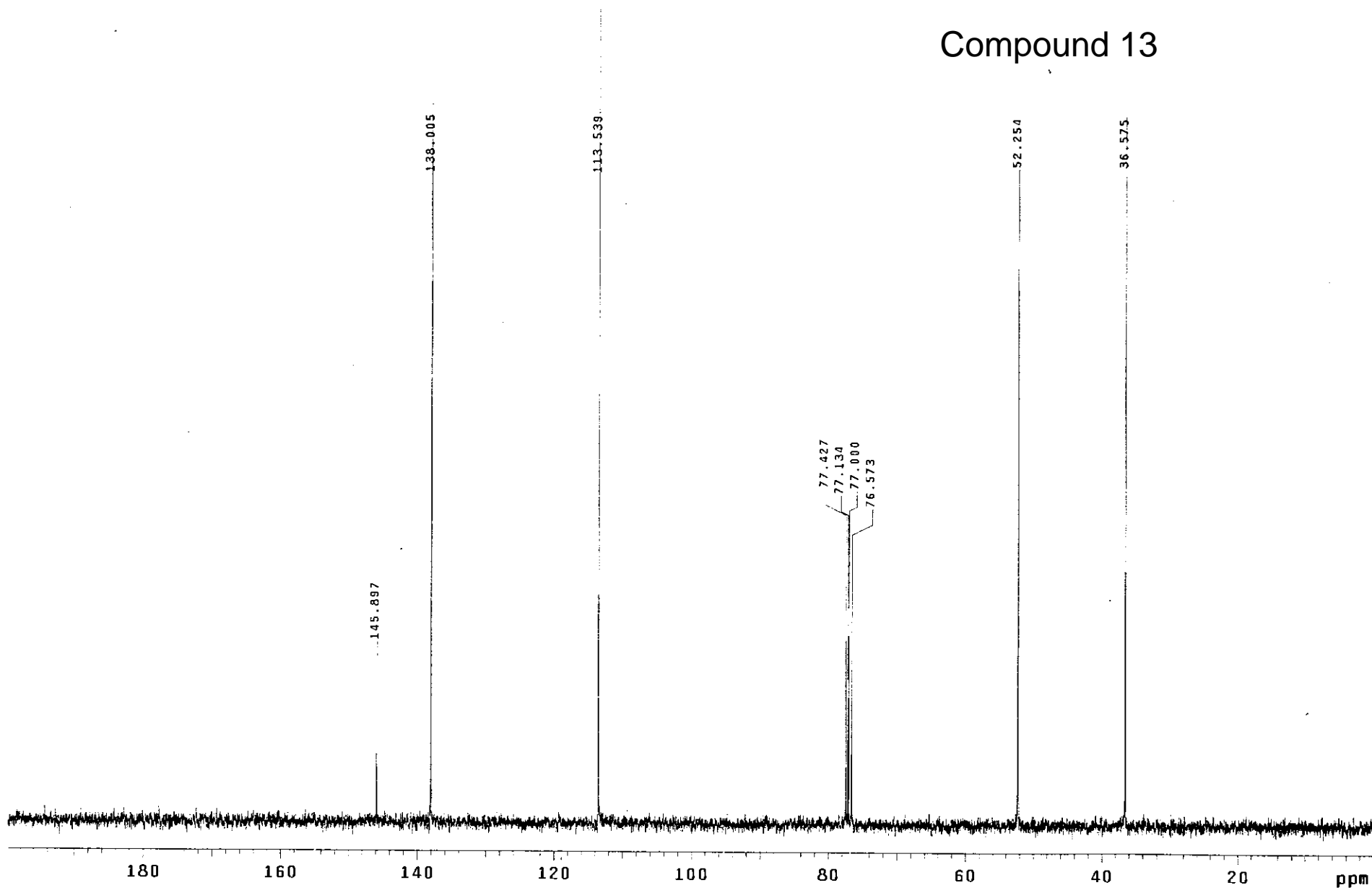
13



q1i011106b

Pulse Sequence: s2pu1

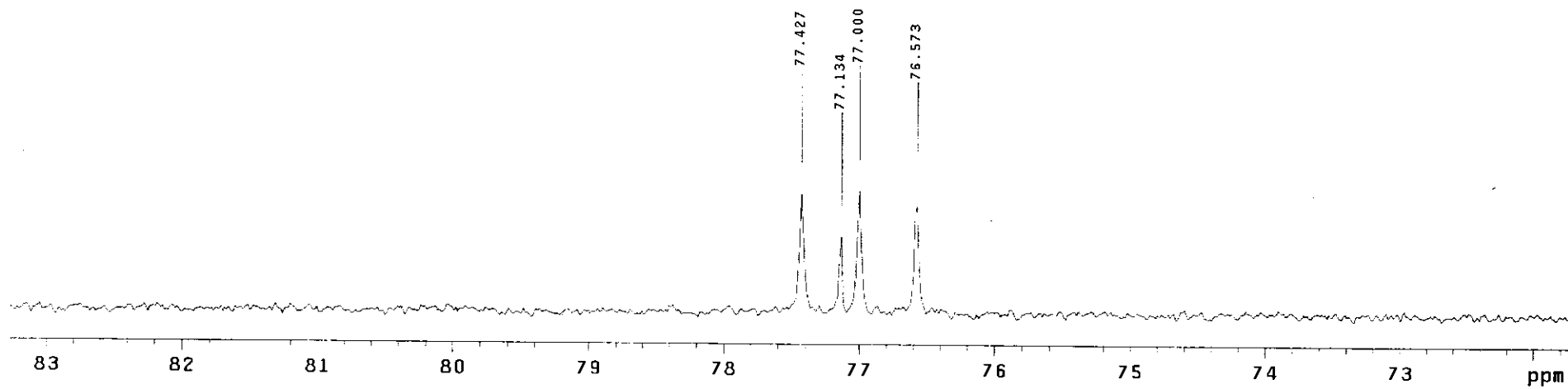
Compound 13



q1i011106b

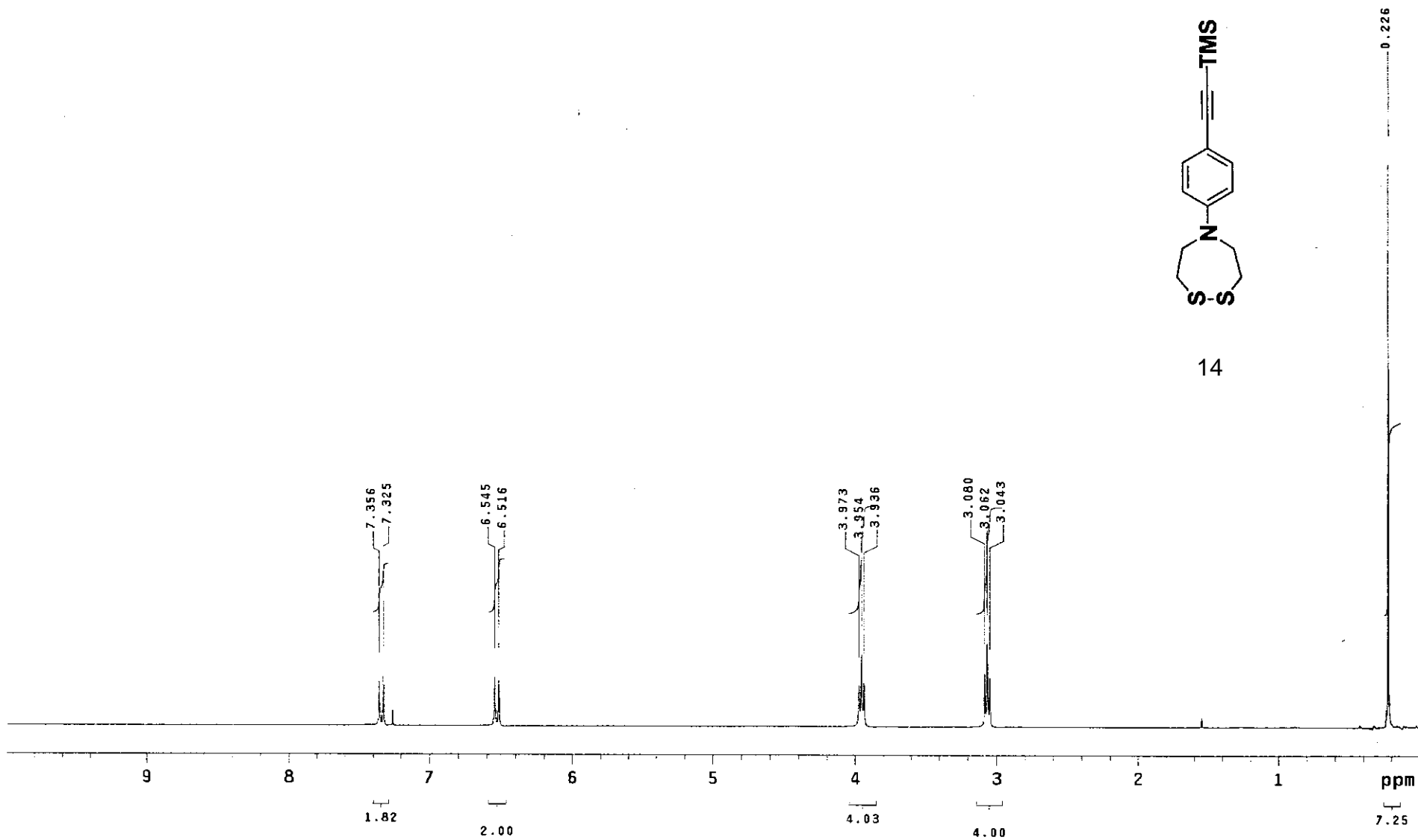
Pulse Sequence: s2pu1

Compound 13



ql1011108a

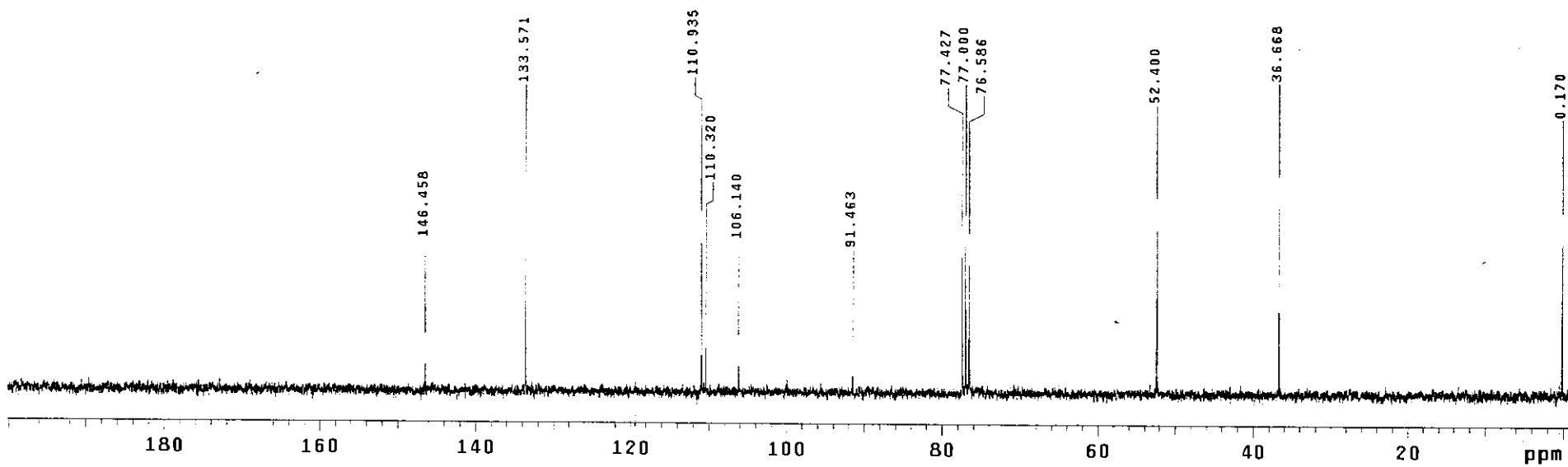
Pulse Sequence: s2pu1



qlf01108b

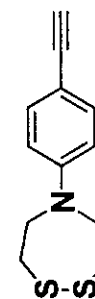
Pulse Sequence: s2pu1

Compound 14

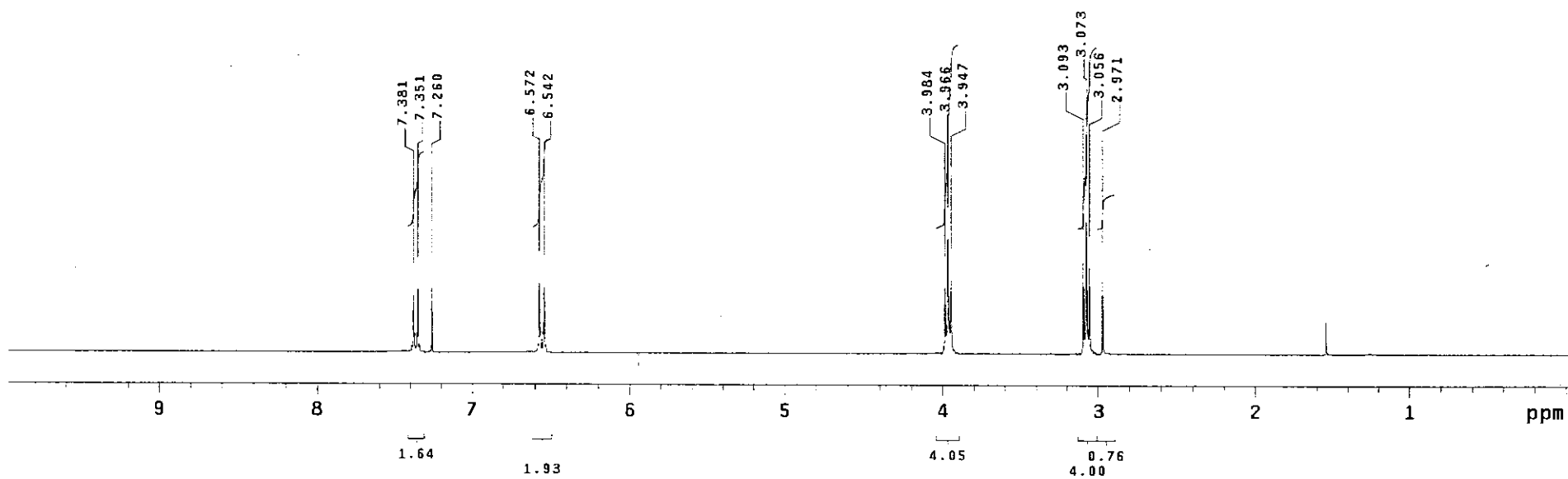


q11011112a

Pulse Sequence: s2pu1



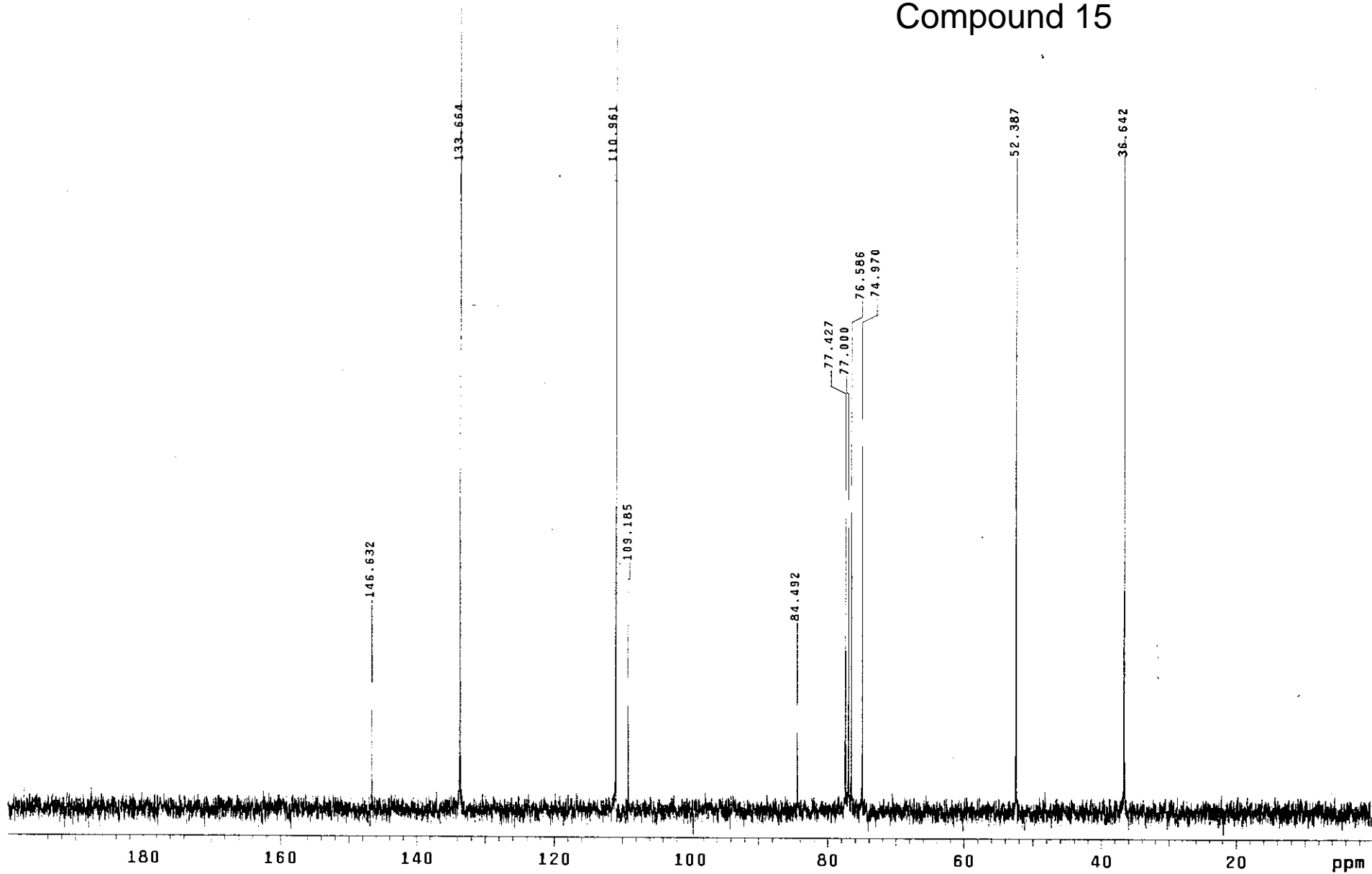
15



q1i011112b

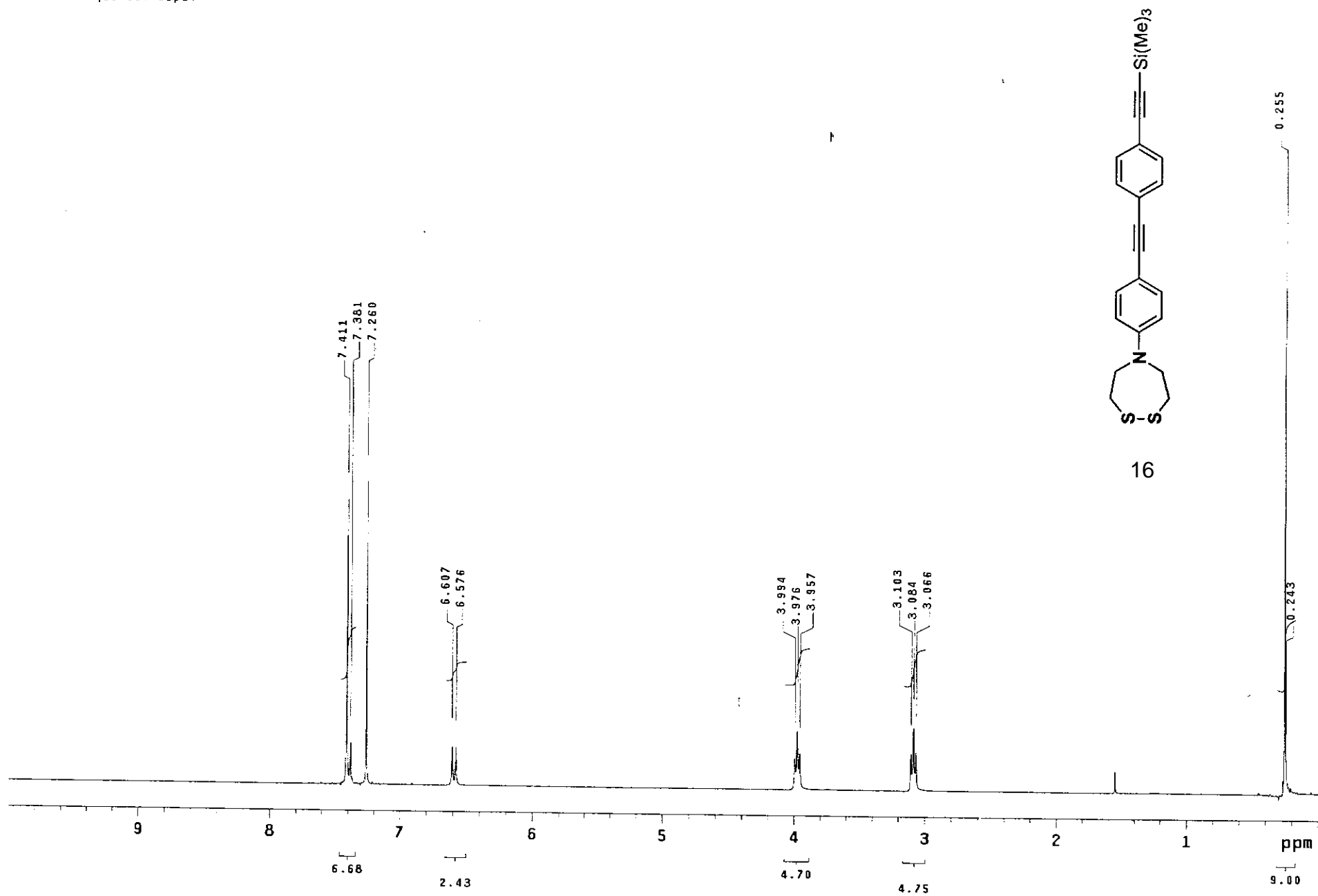
Pulse Sequence: s2pu1

Compound 15



q1i011128a

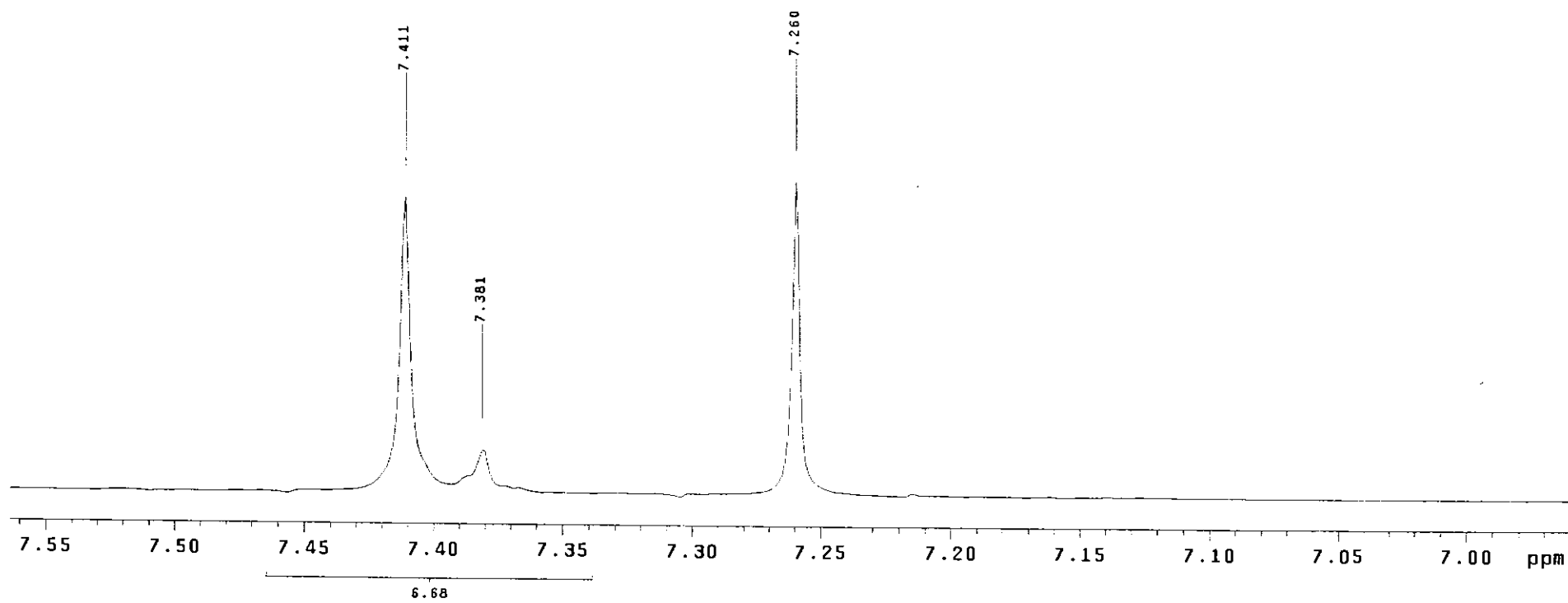
Pulse Sequence: s2pu1



q1i011128a

Pulse Sequence: s2pu1

Compound 16



q11011128

Pulse Sequence: s2pu1

Solvent: CDCl3
Ambient temperature
INOVA-300 "sunofmr"

PULSE SEQUENCE

Relax. delay 1.000 sec
Pulse 42.6 degrees
Acq. time 0.599 sec
Width 16501.7 Hz
500 repetitions

OBSERVE C13, 75.4216991 MHz

DECOUPLE H1, 299.9478514 MHz

Power 32 dB

continuously on
WALTZ-16 modulated

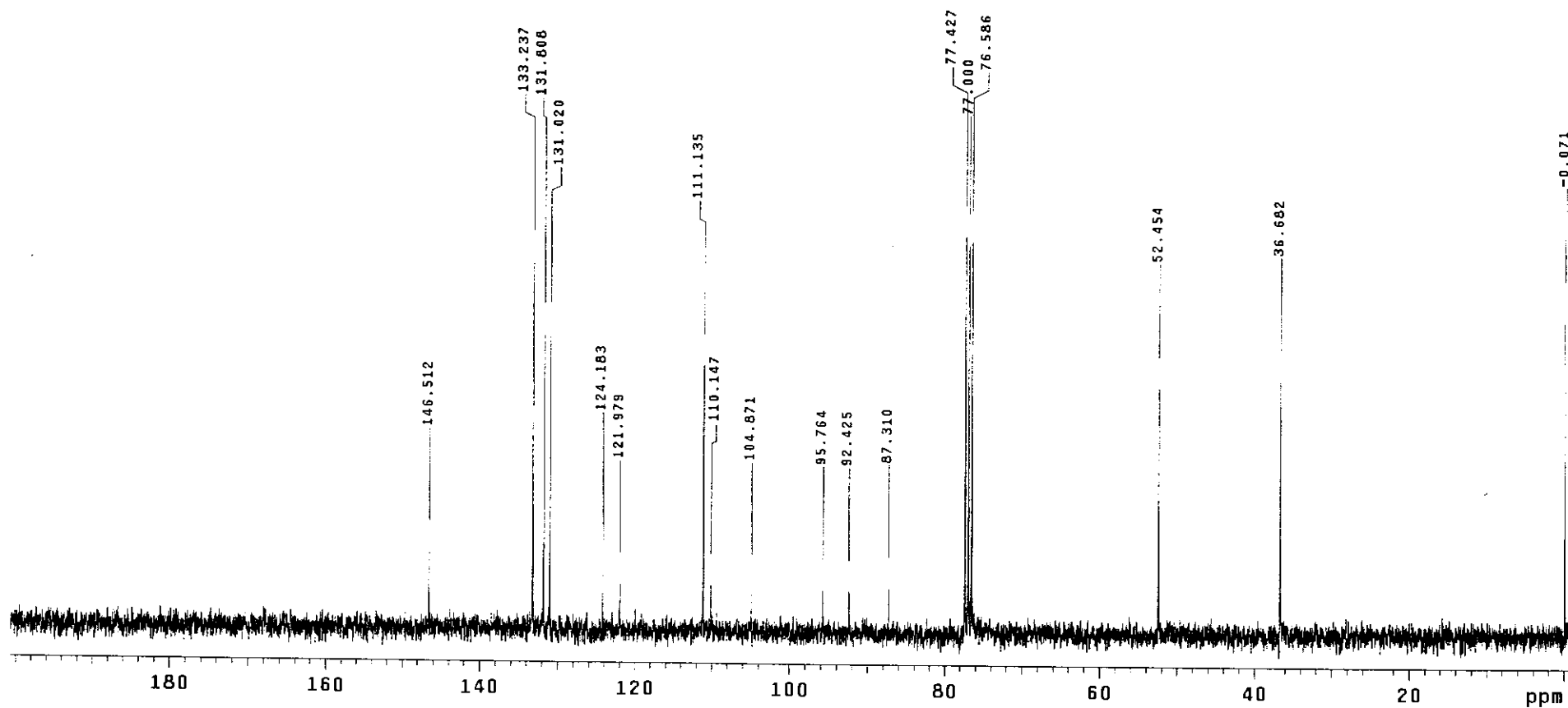
DATA PROCESSING

Line broadening 1.0 Hz

FT size 32768

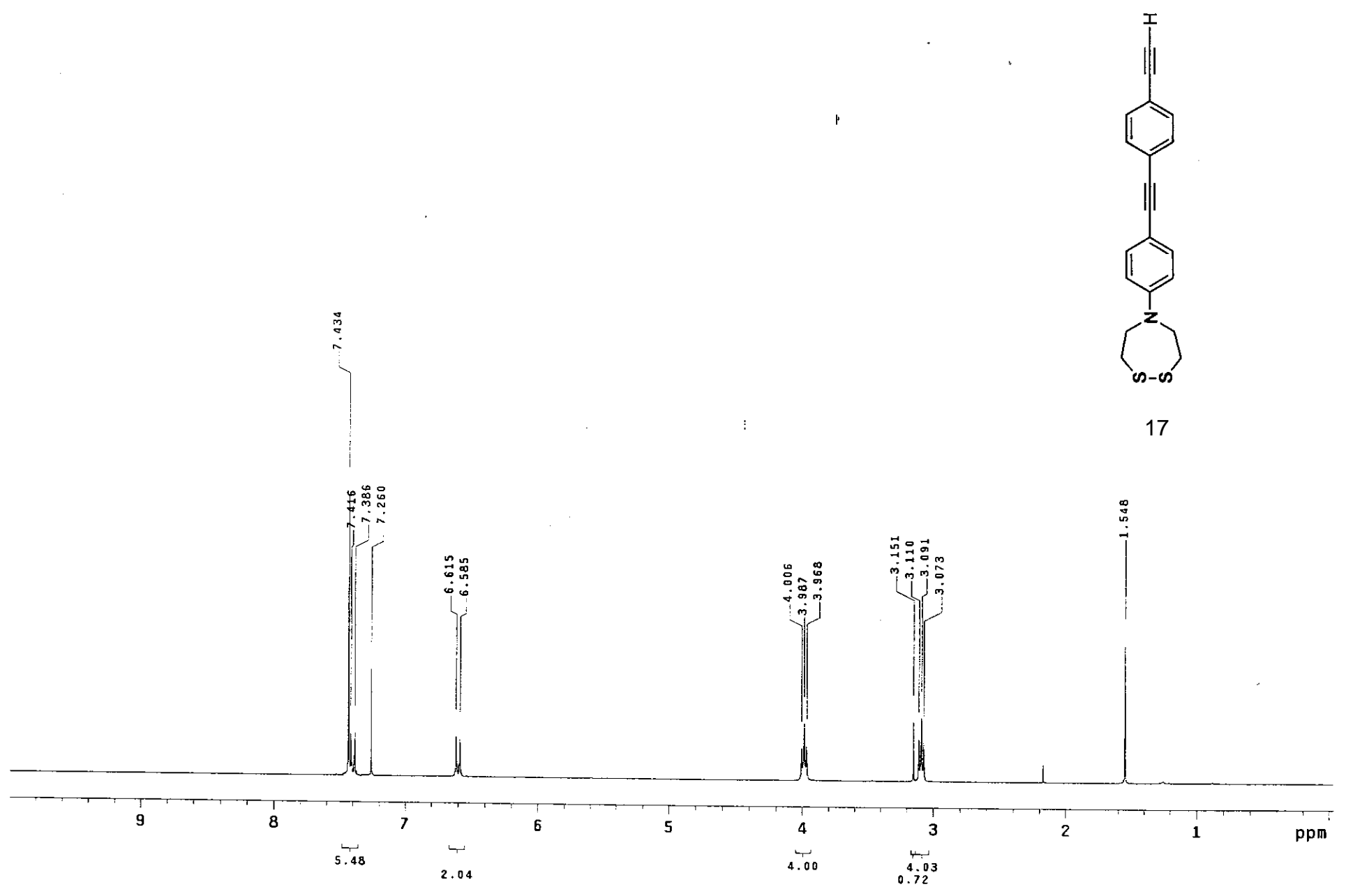
Total time 13 min, 24 sec

Compound 16



q11011204a

Pulse Sequence: s2pu1



17

q1i011204b

Pulse Sequence: s2pu1

Solvent: CDCl3
Ambient temperature
INDVA-300 "sunofnmr"

PULSE SEQUENCE

Relax. delay 1.000 sec

Pulse 42.6 degrees

Acq. time 0.599 sec

Width 16501.7 Hz

300 repetitions

OBSERVE C13, 75.4217001 MHz

DECOUPLE H1, 299.9478514 MHz

Power 32 dB

continuously on

WALTZ-16 modulated

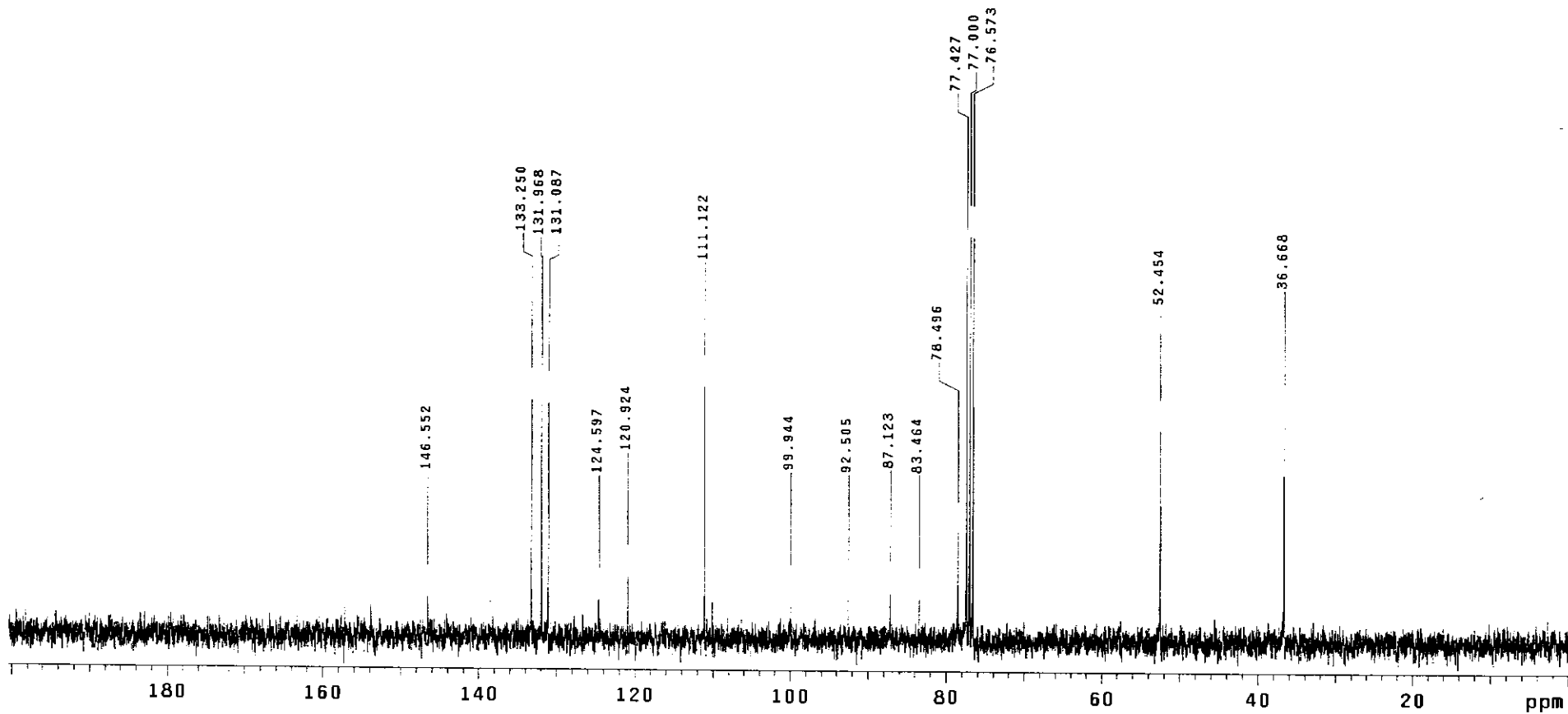
DATA PROCESSING

Line broadening 1.0 Hz

FT size 32768

Total time 8 min, 2 sec

Compound 17



quan11020327b

Pulse Sequence: s2pu1

Solvent: CDC13

Ambient temperature

INOVA-300 "sunofnmr"

PULSE SEQUENCE

Relax. delay 1.000 sec

Pulse 37.9 degrees

Acq. time 2.501 sec

Width 4799.0 Hz

8 repetitions

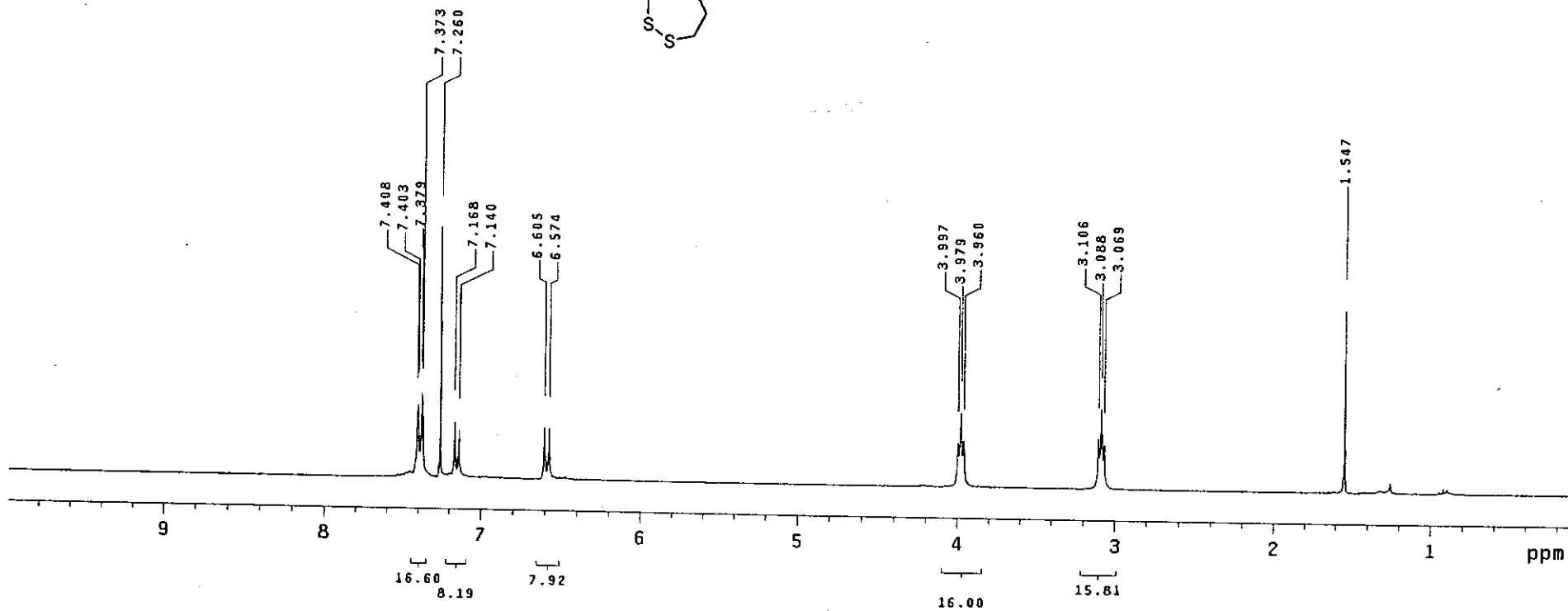
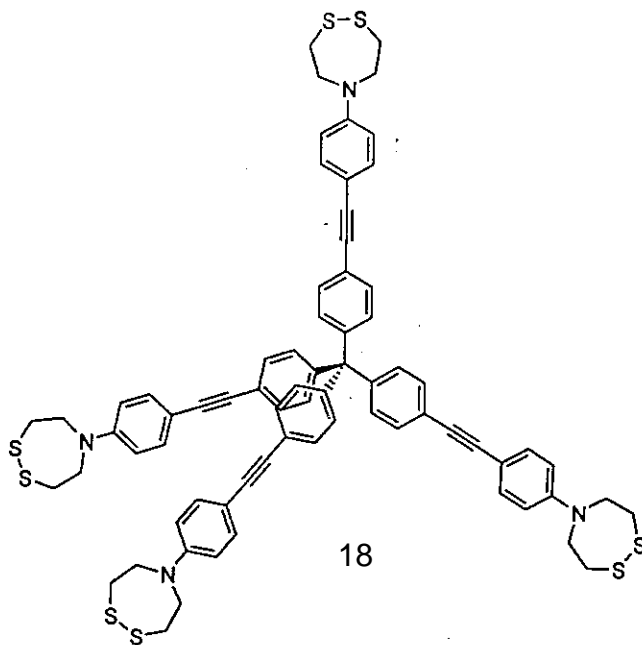
OBSERVE H1, 299.9468615 MHz

DATA PROCESSING

Line broadening 0.2 Hz

FT size 32768

Total time 0 min, 28 sec



quan1020327b

Pulse Sequence: s2pu1

Solvent: CDCl3

Ambient temperature

INOVA-300 "sunofnmr"

PULSE SEQUENCE

Relax. delay 1.000 sec

Pulse 37.9 degrees

Acq. time 2.501 sec

Width 4799.0 Hz

8 repetitions

OBSERVE H1, 299.9468615 MHz

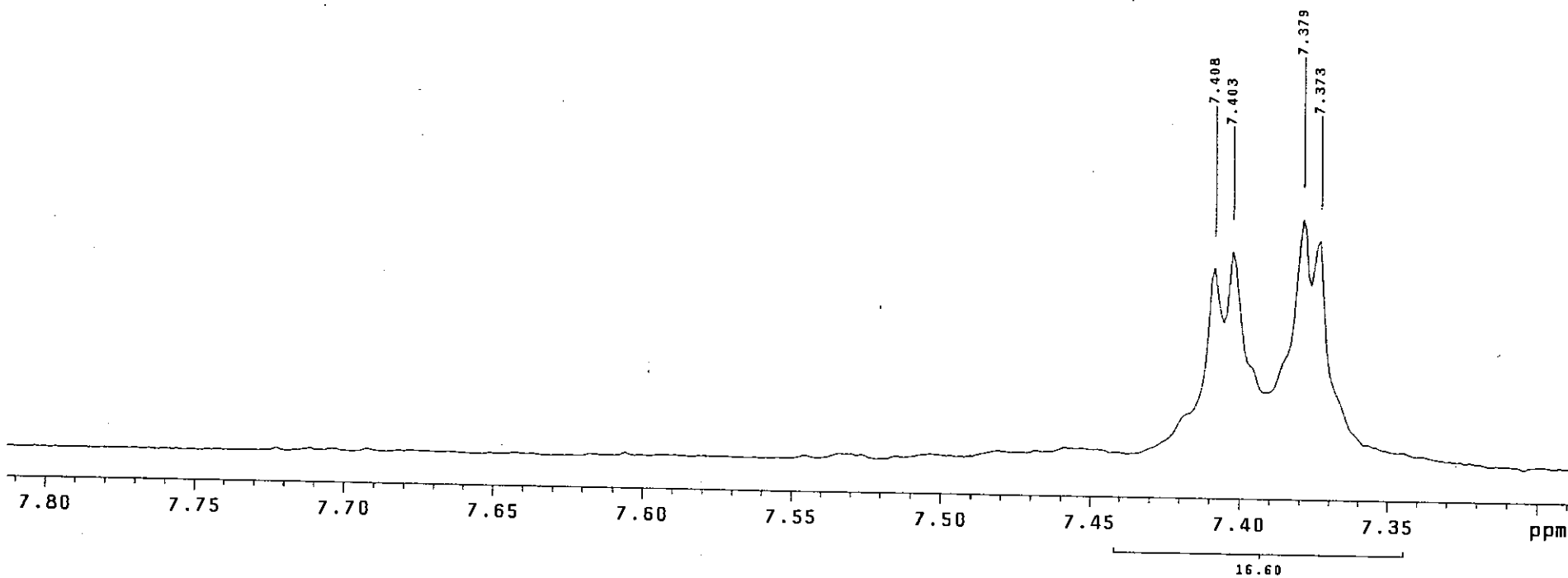
DATA PROCESSING

Line broadening 0.2 Hz

FT size 32768

Total time 0 min, 28 sec

Compound 18



quant1020327b

Pulse Sequence: s2pu1

Solvent: CDCl3
Ambient temperature
INVA-300 "sunofmr"

PULSE SEQUENCE

Relax. delay 1.000 sec
Pulse 37.9 degrees
Acq. time 2.501 sec
Width 4799.0 Hz
8 repetitions

OBSERVE H1, 299.9468615 MHz

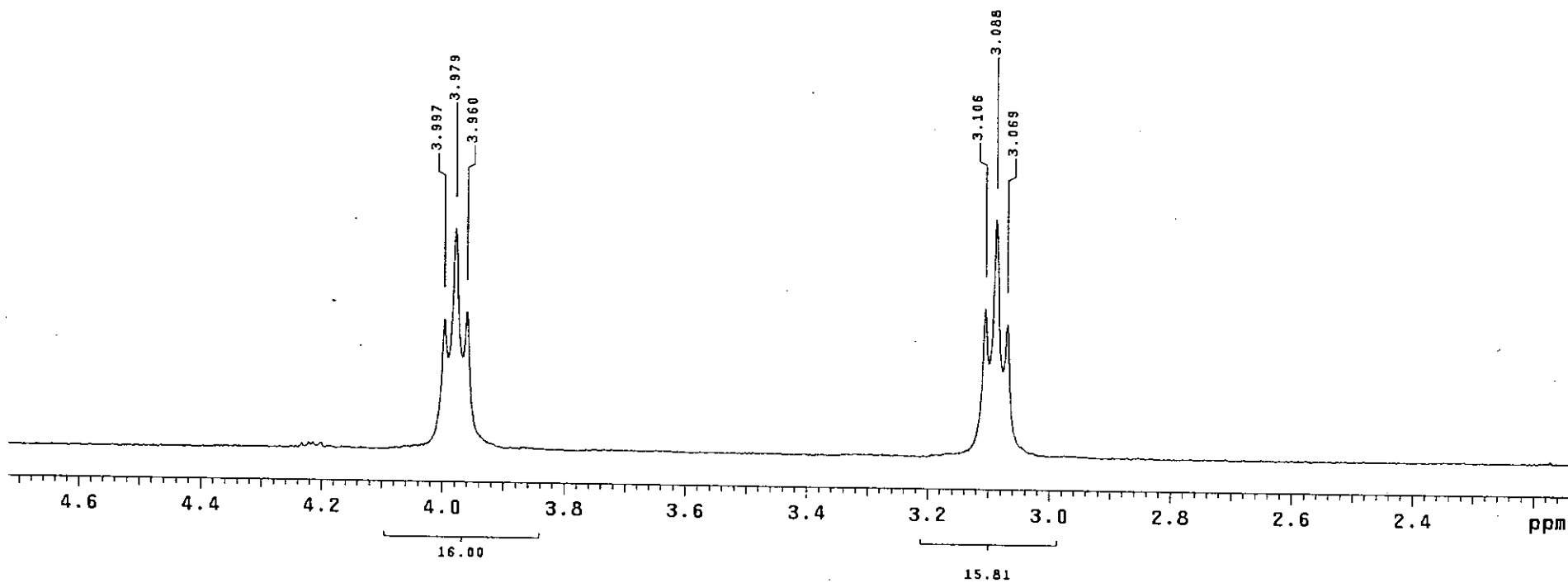
DATA PROCESSING

Line broadening 0.2 Hz

FT size 32768

Total time 0 min, 28 sec

Compound 18



quan1020327d

Pulse Sequence: s2pul

Solvent: CDCl3
Ambient temperature
INOVA-300 "sunofnar"

PULSE SEQUENCE

Relax. delay 1.000 sec
Pulse 42.6 degrees
Acq. time 0.599 sec
Width 16501.7 Hz
10000 repetitions

OBSERVE C13, 75.4216991 MHz
DECOUPLE H1, 299.9478514 MHz

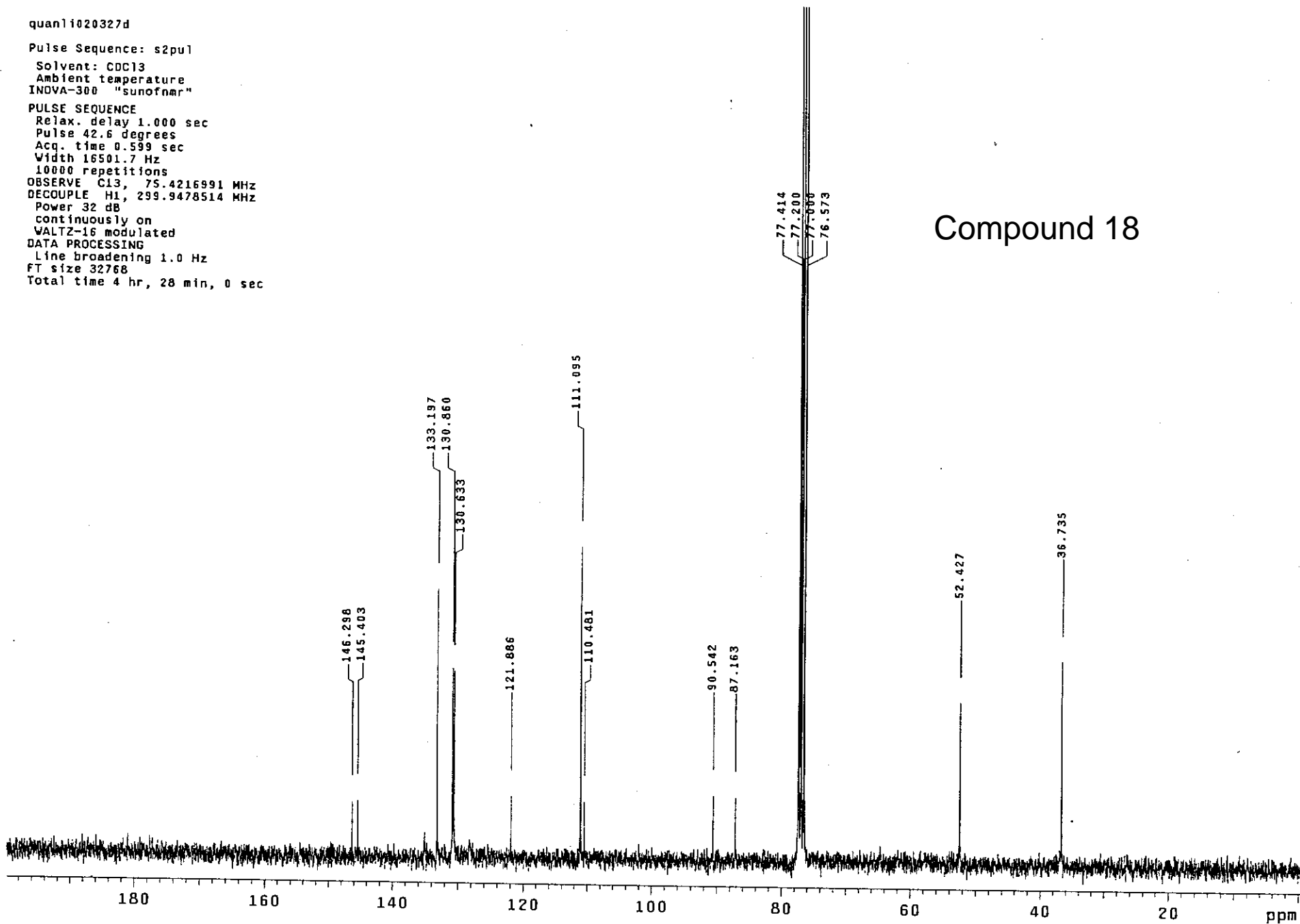
Power 32 dB
continuously on
WALTZ-16 modulated

DATA PROCESSING

Line broadening 1.0 Hz
FT size 32768

Total time 4 hr, 28 min, 0 sec

Compound 18



quan11020327d

Pulse Sequence: s2pu1

Solvent: CDCl3

Ambient temperature

INOVA-300 "sunofnmr"

PULSE SEQUENCE

Relax. delay 1.000 sec

Pulse 42.6 degrees

Acq. time 0.599 sec

Width 16501.7 Hz

10000 repetitions

OBSERVE C13, 75.4216991 MHz

DECOUPLE H1, 299.9478514 MHz

Power 32 dB

continuously on

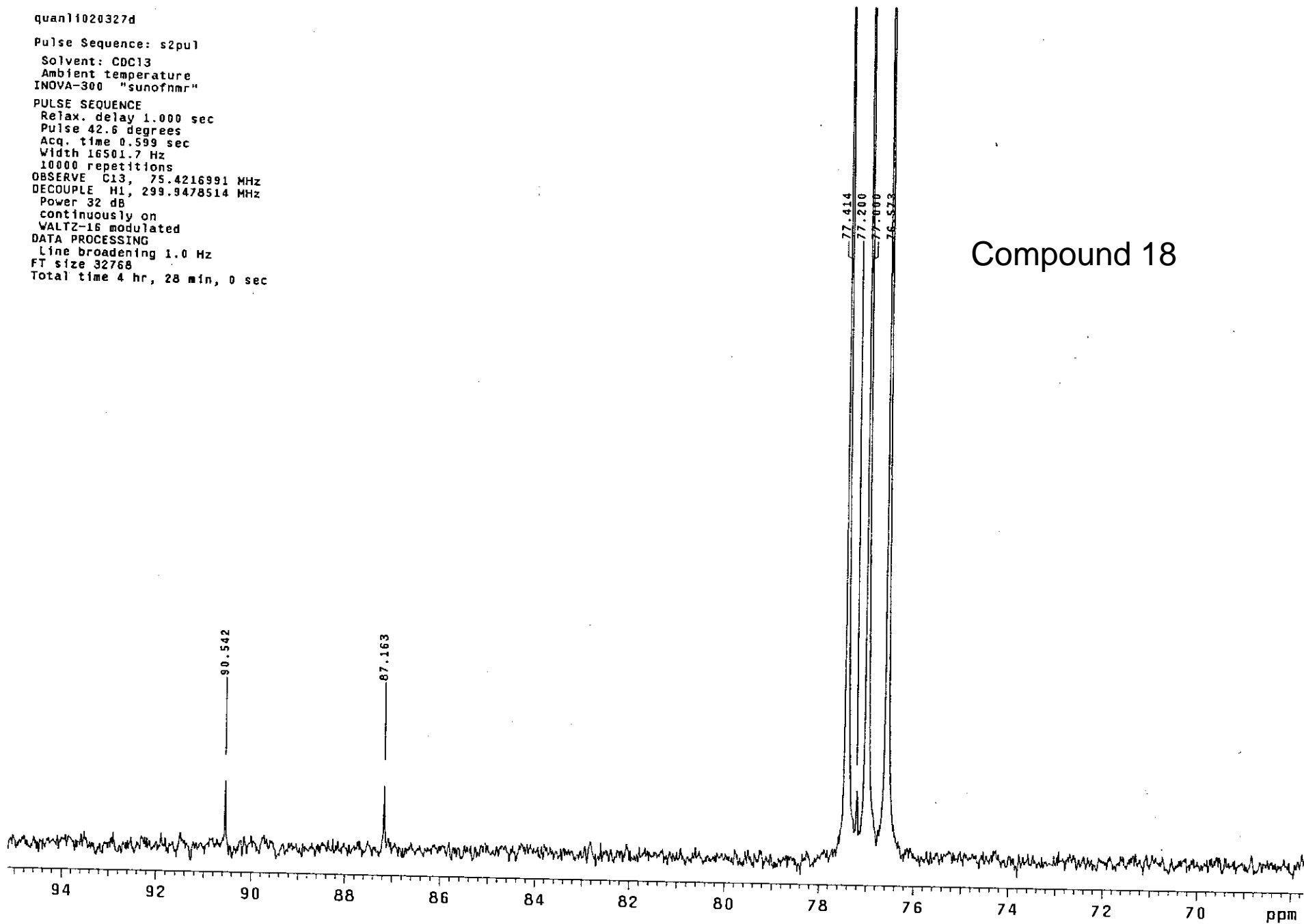
WALTZ-16 modulated

DATA PROCESSING

Line broadening 1.0 Hz

FT size 32768

Total time 4 hr, 28 min, 0 sec



Compound 18

q11011210b

Pulse Sequence: s2pul

Solvent: CDCl3
Ambient temperature
INOVA-300 "sunofnmr"

PULSE SEQUENCE

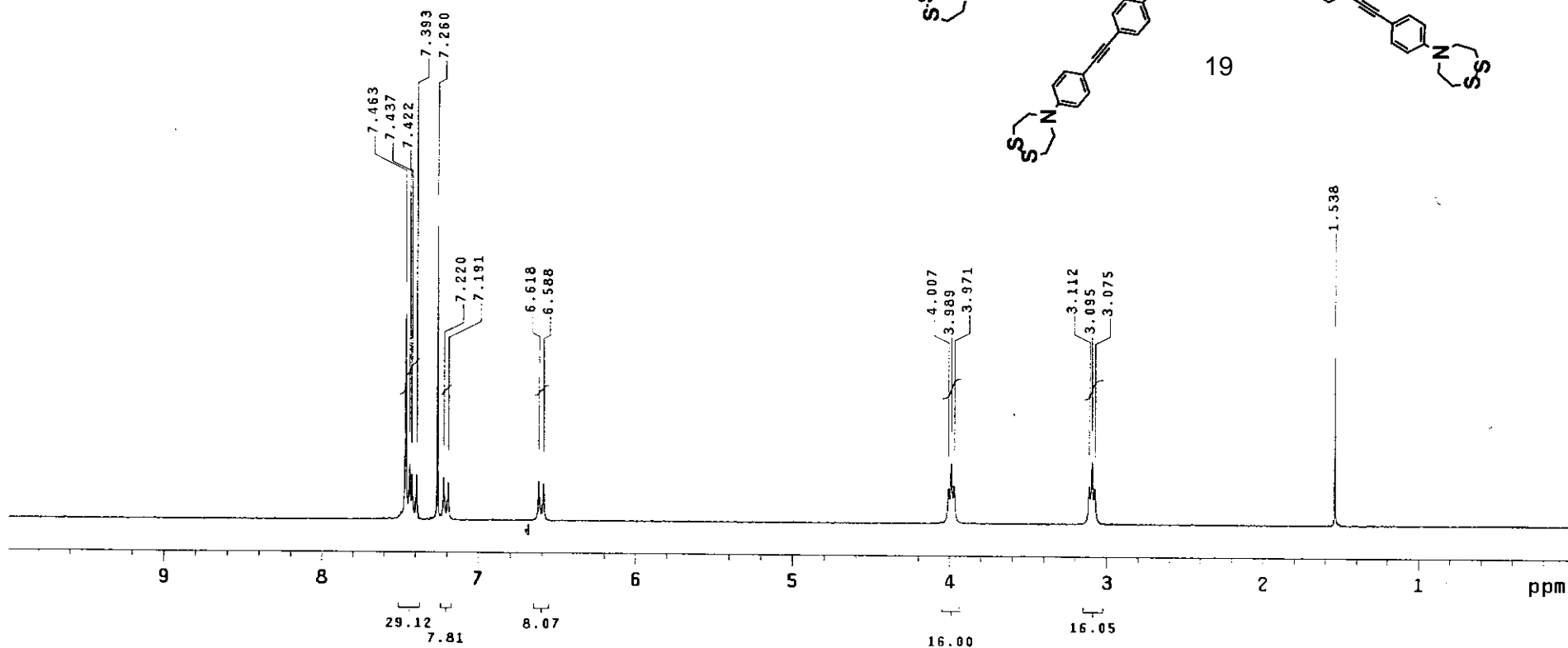
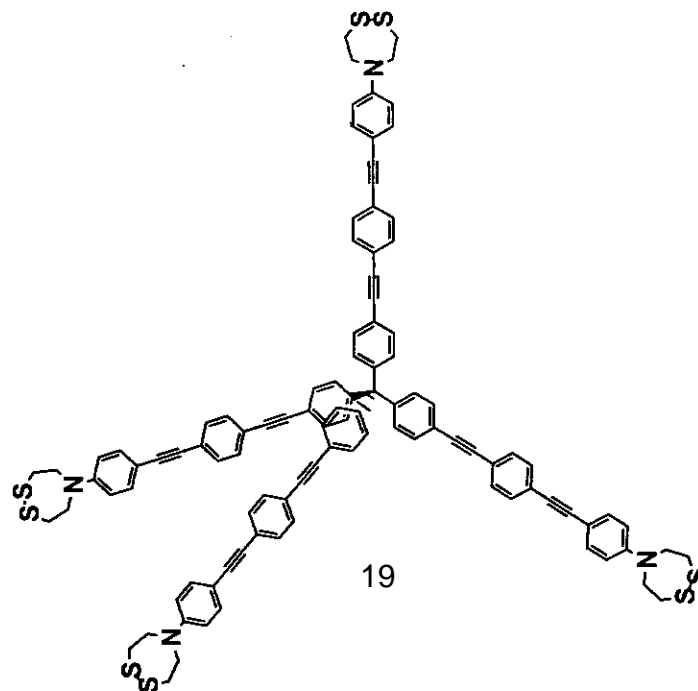
Relax. delay 1.000 sec
Pulse 37.9 degrees
Acq. time 2.501 sec
Width 4799.0 Hz
8 repetitions

OBSERVE H1, 299.9468618 MHz

DATA PROCESSING

Line broadening 0.2 Hz
FT size 32768

Total time 0 min, 28 sec



q11011210b

Pulse Sequence: s2pul

Solvent: CDCl3

Ambient temperature

INOVA-300 "sunofnmr"

PULSE SEQUENCE

Relax. delay 1.000 sec

Pulse 37.9 degrees

Acq. time 2.501 sec

Width 4799.0 Hz

8 repetitions

OBSERVE HI, 299.9468618 MHz

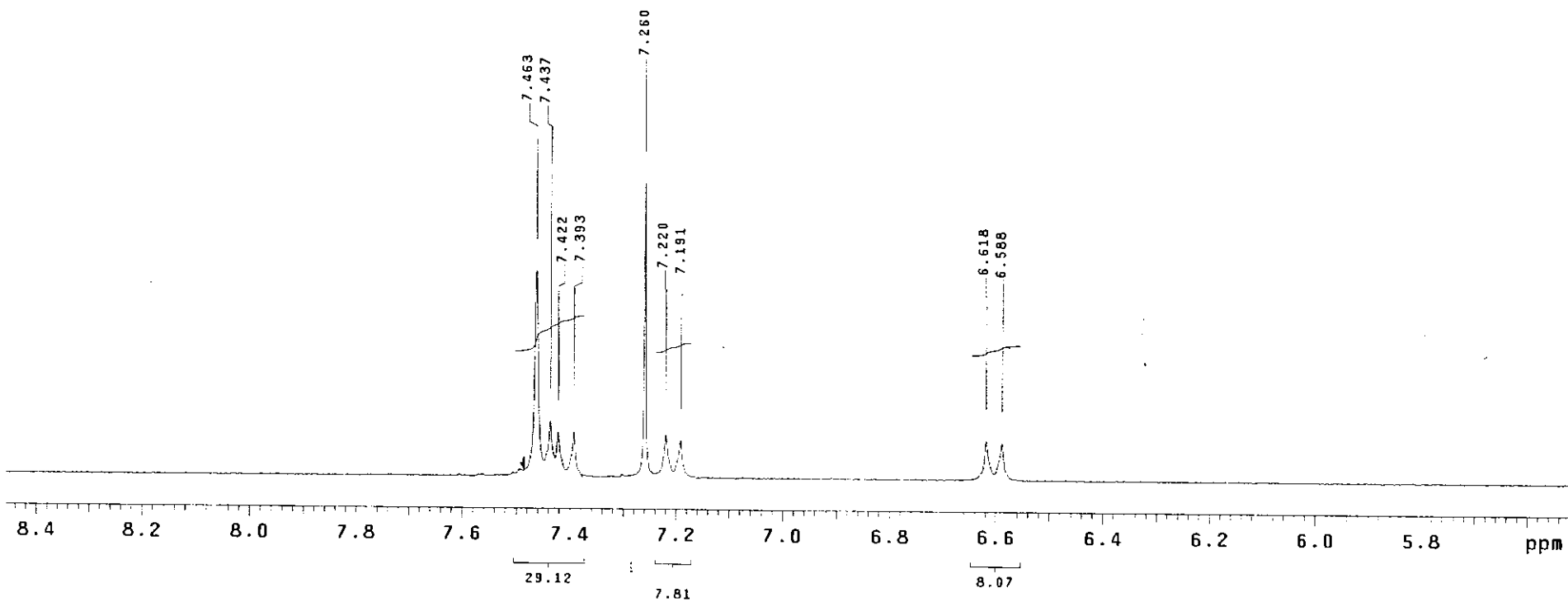
DATA PROCESSING

Line broadening 0.2 Hz

FT size 32768

Total time 0 min, 28 sec

Compound 19



quanlitip19a

Pulse Sequence: s2pu1

Solvent: CDCl3

Ambient temperature

file: quanli020603a

INOVA-300 "sunofmr"

PULSE SEQUENCE

Relax. delay 1.000 sec

Pulse 37.9 degrees

Acq. time 2.501 sec

Width 4799.0 Hz

8 repetitions

OBSERVE H1, 299.9468618 MHz

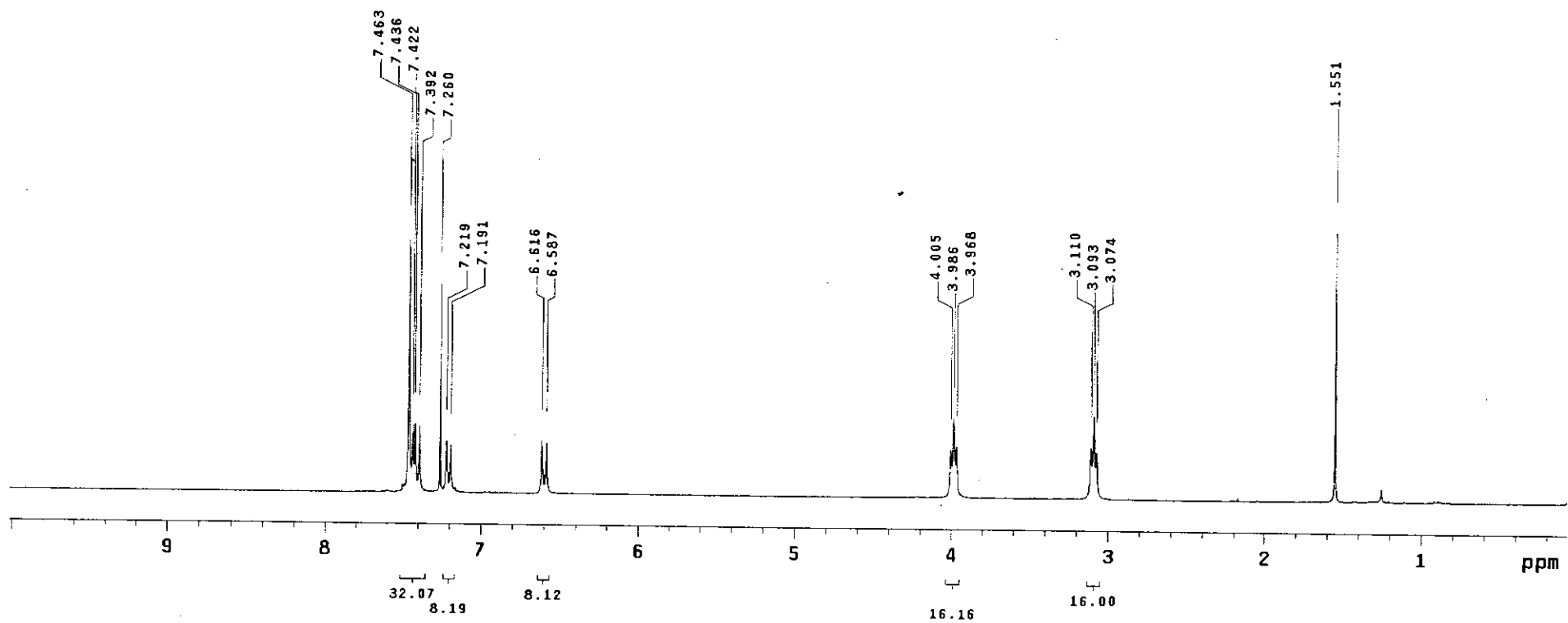
DATA PROCESSING

Line broadening 0.2 Hz

FT size 32768

Total time 0 min, 28 sec

Compound 19



quantitp19a

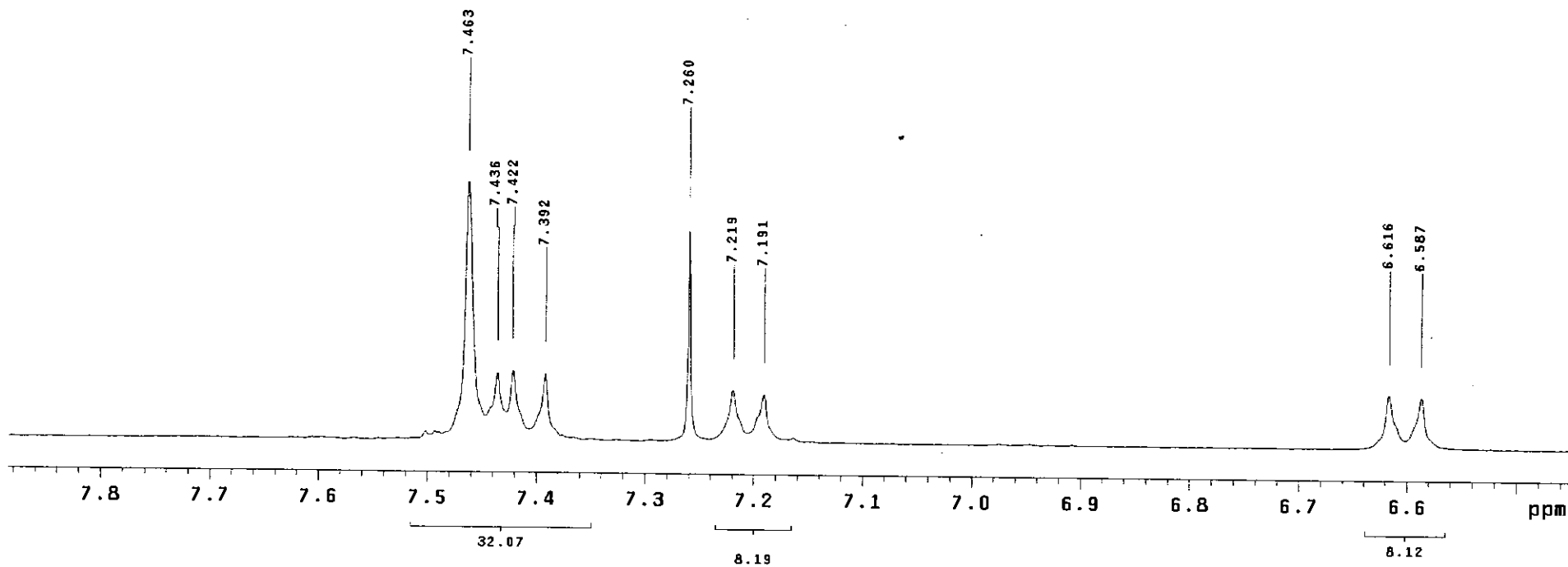
Pulse Sequence: s2pul

Solvent: CDCl3
Ambient temperature
File: quant1020603a
INOVA-300 "sunofnar"

PULSE SEQUENCE

Relax. delay 1.000 sec
Pulse 37.9 degrees
Acq. time 2.501 sec
Width 4799.0 Hz
8 repetitions
OBSERVE H1, 299.9468618 MHz
DATA PROCESSING
Line broadening 0.2 Hz
FT size 32768
Total time 0 min, 28 sec

Compound 19



quan1t1p19

Pulse Sequence: s2pul

Solvent: CDCl3

Ambient temperature

File: quan11020603b

INOVA-300 "sunofnmr"

PULSE SEQUENCE

Relax. delay 1.000 sec

Pulse 42.6 degrees

Acq. time 0.589 sec

Width 16501.7 Hz

25776 repetitions

OBSERVE C13, 75.4216981 MHz

DECOUPLE H1, 299.9478514 MHz

Power 32 dB

continuously on

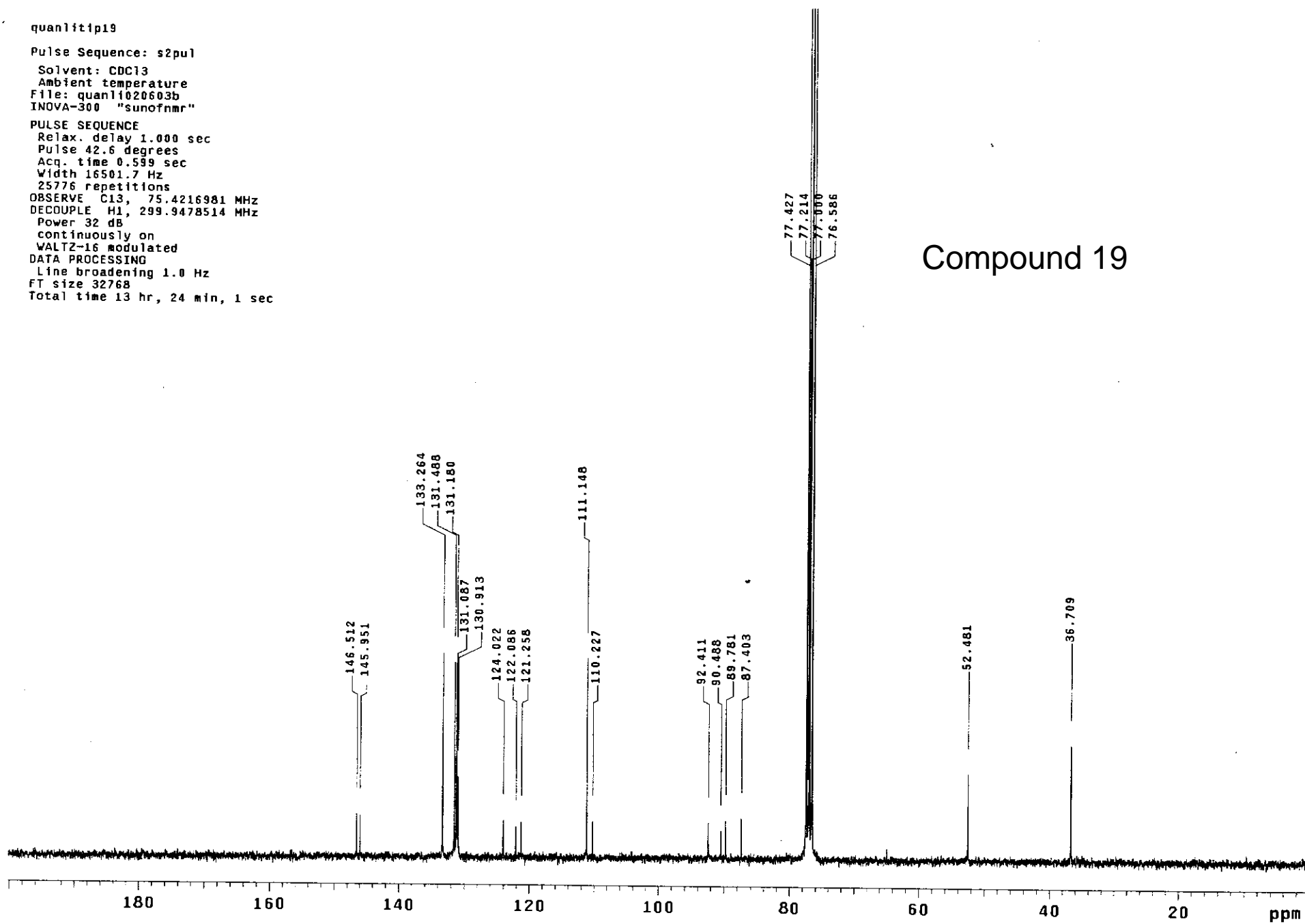
WALTZ-16 modulated

DATA PROCESSING

Line broadening 1.0 Hz

FT size 32768

Total time 13 hr, 24 min, 1 sec



Compound 19

quantitip19

Pulse Sequence: s2pul

Solvent: CDCl3

Ambient temperature

File: quanli020603b

INOVA-300 "sunofnmr"

PULSE SEQUENCE

Relax. delay 1.000 sec

Pulse 42.6 degrees

Acq. time 0.599 sec

Width 16501.7 Hz

25776 repetitions

OBSERVE C13, 75.4216981 MHz

DECOUPLE H1, 299.9478514 MHz

Power 32 dB

continuously on

WALTZ-16 modulated

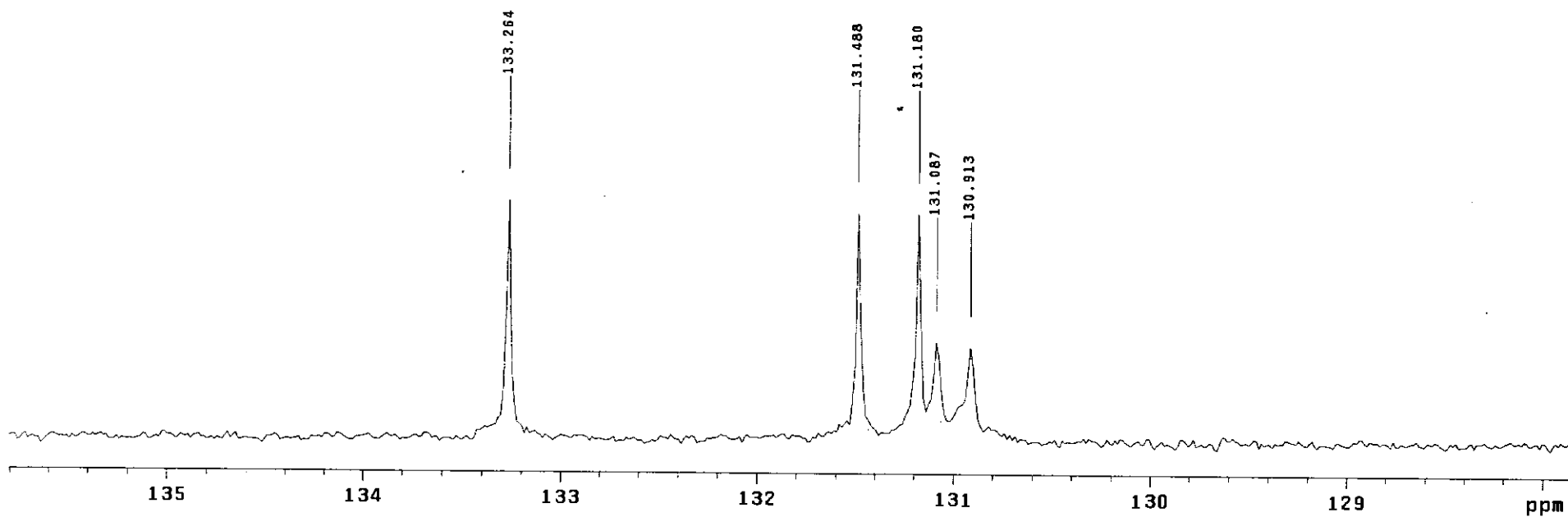
DATA PROCESSING

Line broadening 1.0 Hz

FT size 32768

Total time 13 hr, 24 min, 1 sec

Compound 19



quantitip19

Pulse Sequence: s2pul

Solvent: CDCl3

Ambient temperature

File: quan11020603b

INOVA-300 "sunofnmr"

PULSE SEQUENCE

Relax. delay 1.000 sec

Pulse 42.6 degrees

Acq. time 0.599 sec

Width 16501.7 Hz

25776 repetitions

OBSERVE C13, 75.4216981 MHz

DECOUPLE H1, 299.9478514 MHz

Power 32 dB

continuously on

WALTZ-16 modulated

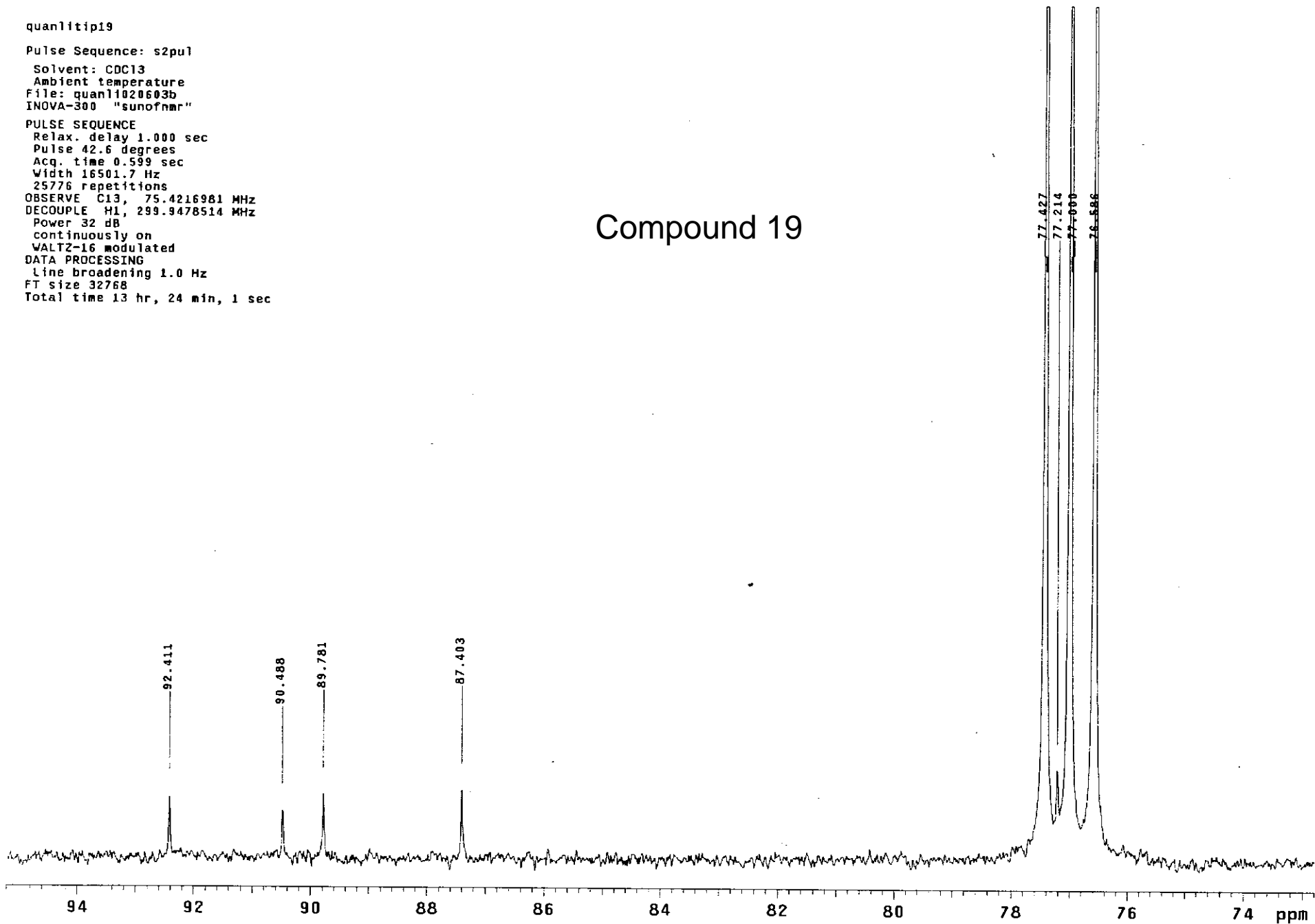
DATA PROCESSING

Line broadening 1.0 Hz

FT size 32768

Total time 13 hr, 24 min, 1 sec

Compound 19



quanli020301spot3a

Pulse Sequence: s2pu1

Solvent: CDC13

Ambient temperature

File: quanli020301spot3a

INOVA-300 "sunofmr"

PULSE SEQUENCE

Relax. delay 1.000 sec

Pulse 37.9 degrees

Acq. time 2.501 sec

Width 4799.0 Hz

8 repetitions

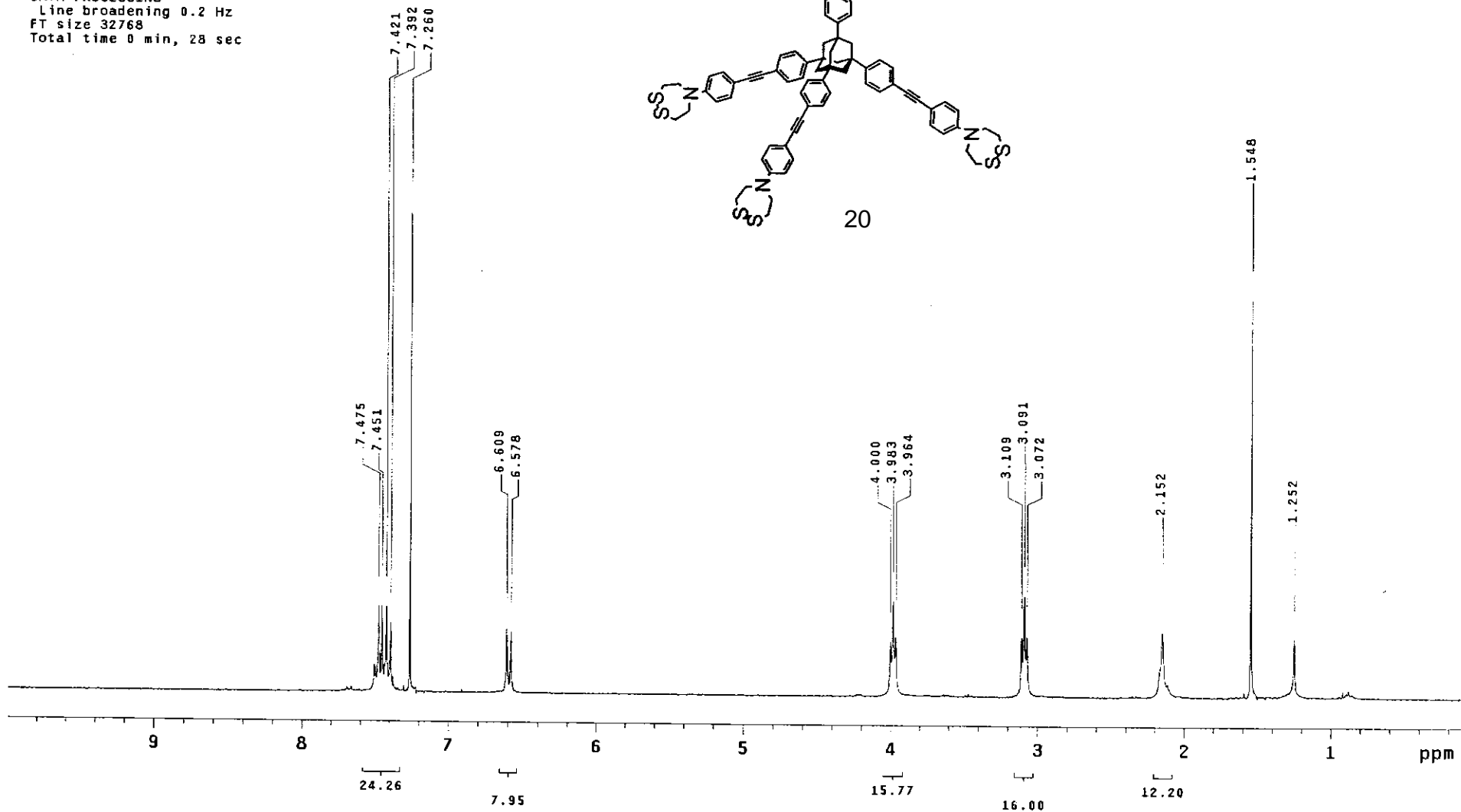
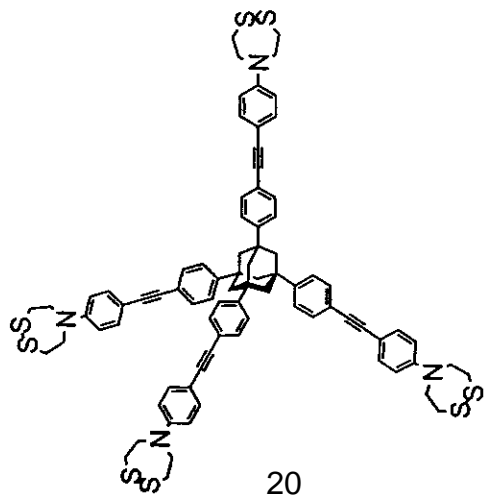
OBSERVE H1, 299.9468615 MHz

DATA PROCESSING

Line broadening 0.2 Hz

FT size 32768

Total time 0 min, 28 sec



quanli020522cT1p

Pulse Sequence: s2pu1

Solvent: CDCl3

Ambient temperature
INOVA-300 "sunofnmr"

PULSE SEQUENCE

Relax. delay 1.000 sec

Pulse 42.6 degrees

Acq. time 0.599 sec

Width 16501.7 Hz

29712 repetitions

OBSERVE C13, 75.4216981 MHz

DECOUPLE H1, 299.9478514 MHz

Power 32 dB

continuously on

WALTZ-16 modulated

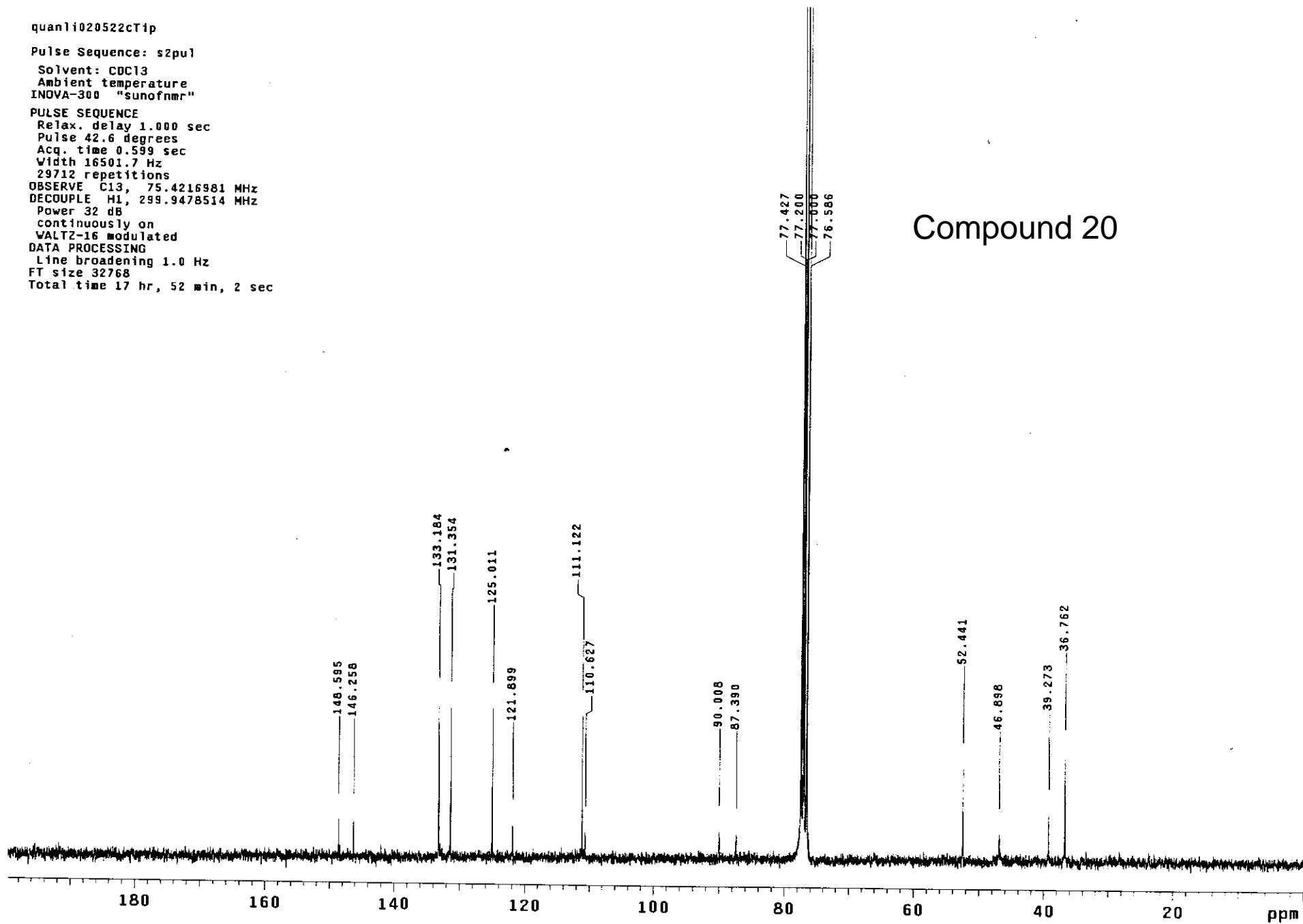
DATA PROCESSING

Line broadening 1.0 Hz

FT size 32768

Total time 17 hr, 52 min, 2 sec

Compound 20



quanli020522cTip

Pulse Sequence: s2pu1

Solvent: CDCl3
Ambient temperature
INOVA-300 "sunofnmr"

PULSE SEQUENCE

Relax. delay 1.000 sec

Pulse 42.6 degrees

Acq. time 0.599 sec

Width 16501.7 Hz

29712 repetitions

OBSERVE C13, 75.4216981 MHz

DECOUPLE H1, 299.9478514 MHz

Power 32 dB

continuously on

WALTZ-16 modulated

DATA PROCESSING

Line broadening 1.0 Hz

FT size 32768

Total time 17 hr, 52 min, 2 sec

Compound 20

